Pleistocene to Holocene planktic foraminiferal biostratigraphy of the Coral Sea, offshore Queensland, Australia.

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Fifteen of 27 potential planktic foraminiferal biostratigraphic events from the latest Pliocene–Holocene have been examined; most of the early Pleistocene was not able to be studied because of a hiatus in the one core which penetrated through the Pleistocene into the top of the Pliocene. Four of these events have been recognised for the first time, and some have been used to establish a formal zonal and subzonal scheme for the region. Two new subzones are proposed, the Globigerinoides quadrilobatus fistulosus Subzone (for the basal part of Zone N.22) and the Bolliella praeadamsi Subzone (for the top part of Zone N.22). These changes have made necessary the redefinition of both the Globorotalia (Truncorotalia) crassaformis viola and the Globorotalia (Truncorotalia) crassaformis hessi Subzones. There is

little evidence that changes in palaeoceanographic conditions are related to the faunal changes observed in the cores, although there are strong indications of reworking at some levels within some of the cores (such as late Miocene Zone N.17 in one core), which may have been related to sea-level fluctuations. The assemblages from the cores show a marked stability, with the faunas being dominated by spinose, oligotrophic taxa. Temperate water forms are either very rare or absent, whereas tropical and subtropical species are dominant. A single new species, Bolliella praeadamsi has been described because of its biostratigraphic utility. This species is an evolutionary intermediate between Bo. calida praecalida and Bo. adamsi.

Introduction

This study is based on a number of cores taken from the eastern Coral Sea over the period December 1981 to December 1987 (Fig. 1). These cores were taken to try to unravel the Pleistocene history of the region and apply the results to the history of the Great Barrier Reef. A planktic foraminiferal study of the cores was made to attempt to establish a biozonation on which palaeoceanographic, stratigraphic palaeotemperature and sea level changes could be correlated. Oxygen isotope stratigraphic studies on one of the cores, 76PC06, by Peerdeman (ANU, personal communication 1990), have enabled some of the biostratigraphic events discussed in this study to be correlated with the oxygen isotope stratigraphic scheme. The zonation covers the interval from the latest Pliocene to the basal Pleistocene (that part of the Matuyama Chron to the top of the Olduvai Subchron), and the interval of the Bruhnes Chron, representing the last 760 000 years (the latest Middle Pleistocene to Holocene, following Berggren & others (1985, p. 227), but the ties to the isotope stratigraphic scheme are restricted to oxygen isotope stage 12 (440 000 yr BP) and younger (Shackleton & Opdyke, 1976).

There have been few attempts to subdivide the Pleistocene in detail using planktic foraminiferids, and these have been summarised in Figure 2. Jenkins (1967) proposed eight subzones for his Globorotalia inflata Zone based on coiling changes in Neogloboquadrina pachyderma for the New Zealand Pleistocene-Holocene. Ericson & Wollin (1968) proposed a biostratigraphic zonation in which the Pleistocene was divided into eight subdivisions based on the presence/ absence of Globorotalia menardii (= Gr. cultrata sensu lato and Gr. tumida sensu lato in this paper) for the Atlantic Ocean Pleistocene. They suggested that the distribution of this species was related to the North American glacial cycles. Blow (1969) subdivided the Pleistocene-Holocene interval into two zones. His N.22, based on the FAD (first appearance datum) of Globorotalia truncatulinoides, spanned the Pleistocene, and N.23, based on the FAD of Globigerinella calida calida, covered the Holocene.

Lamb & Beard (1972) attempted a more detailed biostratigraphic subdivision of the late Neogene, based on material from the Caribbean, Gulf of Mexico and Italian stratotypes, and concentrated mainly on the Pliocene. Two zones were proposed for the Pleistocene–Holocene interval: the Globorotalia truncatulinoides Zone covered the Pleistocene, and the Globorotalia tumida Zone covered the Holocene. The Gr. truncatulinoides Zone was divided into three subzones: a lower Gr. tosaensis Subzone, succeeded by the Globoquadrina dutertrei Subzone, and then by the Pulleniatina finalis Subzone.

Bolli & Premoli Silva (1973) also used material from the Caribbean, but proposed a new subdivision. The Pleistocene-Holocene interval was covered by a single zone, the Globorotalia truncatulinoides truncatulinoides Zone. This zone was subdivided into five subzones: the Pleistocene Globorotalia crassaformis viola Subzone, Globorotalia hessi Subzone, Globigerina calida calida Subzone, and Globigerina bermudezi Subzone, and the Holocene Globorotalia fimbriata Subzone.

Poag & Valentine (1976), in a study of the Louisiana-Texas Basin, proposed a subdivision of the Pleistocene into 12 biostratigraphic and ecostratigraphic zones based mainly on changes in the proportions of Gr. (Globoconella) inflata and Gr. (Gr.) cultrata; the Holocene was recognised by the Gr. (Gr.) ungulata Zone.

In the Red Sea area, Herman (1968) proposed five zones for the last 150 000 years, based on abundance changes in *Gds. quadrilobatus sacculifer* and *Gds. ruber*, as well as the presence or absence of planktic forms. Berggren & Boersma (1969) reached a similar conclusion to that of Herman (1968), based on climatically controlled changes in distribution patterns of planktic species. Reiss & others (1980) slightly modified Herman's (1968) zonation and this was adopted by Locke & Thunell (1988).

Rögl (1974) was the first to apply one of the zonal schemes to sections within the tropical–subtropical Indo-Pacific; he was not able to recognise the two youngest subzones proposed by Bolli & Premoli Silva (1973) in the Timor Sea area because of the absence of the nominate taxa (which are apparently restricted to the Caribbean area).

Thunell (1984) summarised the problems associated with Pleistocene planktic foraminiferal biostratigraphy. He noted that there were two types of zonal scheme. Some used qualitative criteria such as presence/absence data (e.g. the distribution of *Globorotalia cultrata* group; Ericson & Wollin, 1968), or the first or last appearance datums used in most schemes. Others used quantitative criteria such as single-

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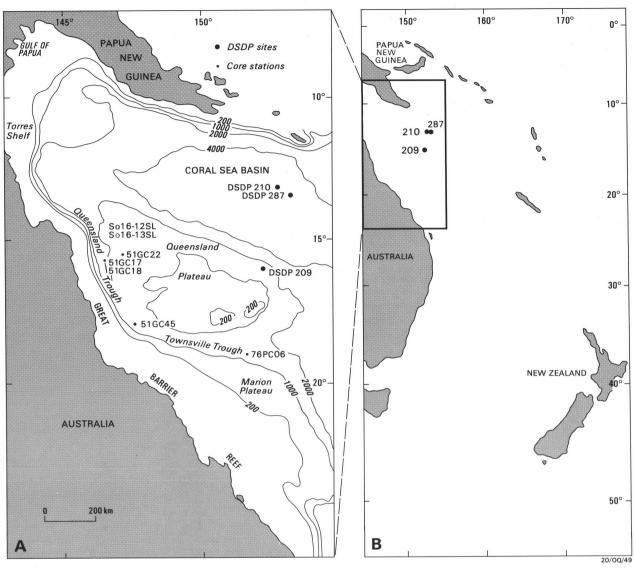


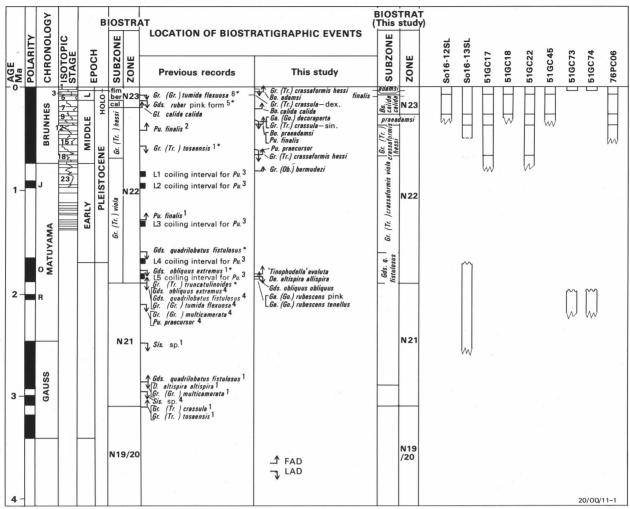
Figure 1. Locality map showing locations of cores used in this study.

species abundances (e.g. Herman, 1968; Thunell & others, 1977), coiling ratios (e.g. Kennett, 1976), or relative abundances of species in the total planktic assemblage (e.g. Kennett & Huddlestun, 1972). Qualitative schemes such as those of Ericson & Wollin (1968) are very useful in the Atlantic Ocean Pleistocene, especially in areas where temperature changes had maximum impact on the faunas. This distribution of the *Gr. cultrata* group is not observed in the Indo-Pacific region (Thunell, 1984). Similarly, most of the quantitative schemes also rely on temperature-linked effects (such as coiling changes in *Neogloboquadrina pachyderma*; see Jenkins, 1967), and are best for regions where temperature variations have been greatest.

All the biostratigraphic zonations discussed above, except Blow (1969) and Rögl (1974), are based on sections deposited either in temperate areas or near the subtropical–temperate transition; that of Blow (1969) is based on sections from the subtropical–tropical Atlantic, whilst that of Rögl (1974) is from the eastern tropical Indian Ocean. The maximum impact of temperature and oceanographic changes on faunal assemblages during the Pleistocene–Holocene interval was probably in boundary areas between climatic zones; changes within these faunal assemblages were due mainly to migratory rather than evolutionary events. Thus the greatest problem for the use of these schemes in warm water areas is the potential absence of

key taxa due to their often restricted geographic ranges; sealevel changes and climatic factors are less marked in these areas, and so have little effect on fossil assemblages. Such climatic and oceanographic effects would be expected to have most effect on faunas in the transitional areas between climatic belts.

The Pleistocene-Pliocene boundary is no longer believed to be equivalent to the Zone N.21/N.22 boundary, as the marker event, the FAD of Gr. (Tr.) truncatulinoides, took place within the latest Pliocene (Rio & others, 1984a,b). Furthermore, this FAD is diachronous, occurring at older stratigraphic levels in higher latitudes and being oldest in the southwest Pacific area (Dowsett, 1989; Hills & Thierstein, 1989). In spite of this, biostratigraphic evidence suggests that the age of this event is consistent for global low latitudes and the use of this FAD to mark the N.21/22 boundary (Blow's (1969) criterion for the base of Zone N.22) is therefore retained for this study. Though the FAD of Gr. (Tr.) truncatulinoides was once used to mark the Pliocene-Pleistocene boundary (Blow, 1969; Kennett & Srinivasan, 1983), Rio & others (1984a,b) have shown that this event occurs well below the boundary in most Mediterranean sections; the FAD of Globigerina (Globigerina) cariacoensis now marks this boundary in the Mediterranean area. However, as this species has not been recorded from the tropical Pacific, the LAD (last appearance datum) of Gds. quadrilobatus



1 Berggren et al 1985 2 Lamb & Beard 1975 3 Saito 1976 4 Kennett & Srinivasan, 1983 5 Thompson et al 1979 6 Bolli & Premoli Silva, 1973 *Previous records used in this study.

Figure 2. Biostratigraphic summary for nine gravity and piston cores taken in the Queensland and Townsville Troughs. Timescale after Bolli & Premoli Silva (1973) and Chaproniere (1985b); oxygen isotope scale after Shackleton & Opdyke (1976).

fistulosus appears to be the closest planktic foraminiferal event to the Pliocene–Pleistocene boundary, and is probably the most suitable to mark this boundary; this event has been dated at 1.6 Ma by Berggren & others (1985).

The biostratigraphic scheme proposed in this paper is based on both qualitative and quantitative criteria. Though FADs and LADs of the various morphotypes are the main events used, changes in coiling direction are important. Presence/absence data, changes in single species and relative species abundances have not been used, due to the faunal stability (the regional temperatures probably did not fall below those of the tropical belt during cooler periods elsewhere).

A reconnaissance oxygen isotope study was made by F. Peerdeman (ANU) on samples taken at 15 cm intervals on core 76PC06. The results of this study have been used here. Peerdeman's study shows that part of the top of the core was lost during coring operations (which is supported by the planktic foraminiferal studies). This, together with the presence of turbidites (indicated by the values of $\delta^{18}O$) makes evaluation of the oxygen isotope stage uncertain. In addition, levels below 700 cm in the core are less reliable for oxygen isotope stage correlation.

Plates 1 to 4 illustrate the key biostratigraphic species as well as other biostratigraphically important forms. All type and figured

specimens are housed in the Commonwealth Palaeontological Collection of the Bureau of Mineral Resources, Canberra, with numbers prefixed by CPC.

Sections studied

Piston and gravity cores collected during Cruise So16 of the R/V Sonne, and Cruises 51 and 76 of the R/V Rig Seismic in the southeastern margin of the Coral Sea Basin, form the basis for this study. The locations of these cores are given in Figure 1. Reworked faunas from the Late Miocene or Pliocene, mainly found in turbidites, are present in some of the cores.

All but one of the cores bottomed in middle Pleistocene sediments. Core So16-13SL bottomed in the late Pliocene (Matuyama Chron), but all the lower Pleistocene (that is, the section from the top of the Olduvai Subchron to the lower part of the Bruhnes Chron, following Berggren & others, 1985) is absent. Five cores (51GC17, 51GC18, 51GC22, 51GC43 and 76PC06) penetrated into the middle Pleistocene.

Planktic foraminiferal succession

The distribution of selected biostratigraphically important taxa is discussed below for each core separately. The distribution of all planktic taxa recorded from each core is given in Figures 3 to 0.

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1166
640017-000
640018-025
640020-050
640021-075
640022-100
640023-125
640024-150
640026-175
640027-200
640028-225
                              Bo. adamsi
                              Bo. calida calida
                              Bo. calida praecalida
                              Bo. praeadamsi
                              Ca. nitida nitida
                              De.
                                   conglomerata
                                   (Be.) digitata digitata
(Be.) digitata praedigitata
(Ga.) bulloides bulloides
                              Ga.
                              Ga.
                              Ga.
                                   (Ga.)
(Ga.)
                                          bulloides umbilicata
                           .
                              Ga.
                              Ga.
                                          falconensis
                                   (Ga.) pseudofoliata
                              Ga.
                                   (Ga.) quinqueloba
(Go.) rubescens rubescens—pink
                              Ga.
                              Ga.
                                   (Go.) rubescens rubescens—white (Go.) rubescens tenellus
                              Ga.
                              Ga.
                              GI.
                                  aequilateralis
                                  glutinata glutinata
                              Gt.
                              Gt.
                                  glutinata naparimaensis
                              Gt.
                                   glutinata parkerae
                              Gt. uvula
                              Gds. conglobatus conglobatus
                                     elongatus
                              Gds.
                                     quadrilobatus ?fistulosus
                              Gds.
                                     quadrilobatus sacculifer
                              Gds.
                              Gds. ruber pink form
                                     ruber white form
                              Gds.
                                   (Gb.)
(Gb.)
                                          crassaconica
                              Gr.
                              Gr.
                                          inflata
                                          triangula
                              Gr.
                                   (Gb.)
                              Gr.
                                   (Gr.)
                                          cultrata cultrata
                                   (Gr.)
                                          cultrata fimbriata
                              Gr.
                              Gr.
                                          cultrata menardii
                                   (Gr.)
                              Gr.
                                   (Gr.)
                                          cultrata neoflexuosa
                                   (Gr.)
                                          tumida flexuosa
                              Gr.
                                   (Gr.)
(Gr.)
                              Gr.
                                          tumida tumida
                              Gr.
                                          ungulata
                                   (Ob.) bermudezi
                              Gr.
                                   (ОЬ.)
(ОЬ.)
                                          hirsuta hirsuta
scitula scitula
                              Gr.
                              Gr.
                              Gr.
                                   (Tr.)
                                         crassaformis crassaformis
                                   (Tr.)
(Tr.)
                              Gr.
                                          crassaformis hessi
                                         crassaformis ronda
                              Gr.
                              Gr.
                                   (Tr.)
                                         crassaformis viola
                                   (Tr.)
(Tr.)
                              Gr.
                                         crassula dextral
                                         crassula sinistral
                              Gr.
                                         truncatulinoides pachytheca
truncatulinoides truncatulinoides
                                   (Tr.)
(Tr.)
                              Gr.
                              Gr.
                              H. pelagica
                              Ng. dutertrei
                              Or. universa
                              Or. universa bilobate form
                              Pa. pseudopima
                              Pu.
                                   finalis
                              Pu.
                                   obliquiloculata
                              Sp.
Sp.
St.
                                   dehiscens dehiscens
                                    dehiscens excavata
                                   tokelauae
                              Te.
                                   anfracta
                           .
                              Te.
                                   cf. minutissima
                              Te.
                                   iota
                              Ti.
                                    evoluta
              .
                               Tu. humilis
-0000
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-1750
-1225
-242
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Figure 3. Biostratigraphic range chart for core So16-12SL.

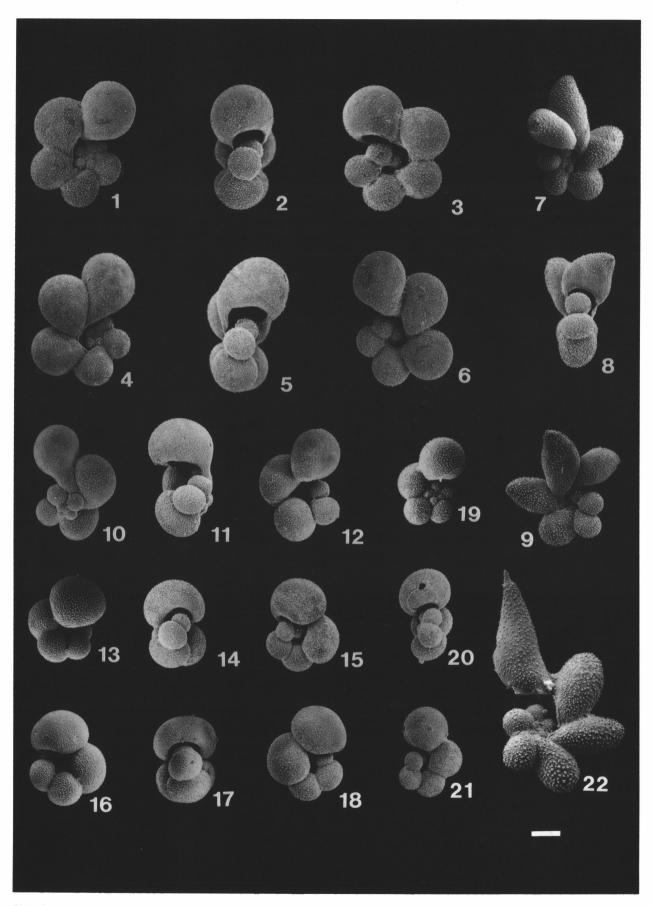


Plate 1. 1–3, Bolliella praeadamsi n. sp., holotype, CPC30216. 4–6, 19–21, Bolliella praeadamsi n. sp., paratypes, CPC30217, CPC30218. 7–9, 22, Bolliella adamsi, CPC30219, CPC30220. 10–12, Bolliella calida calida, CPC30221. 13–18, Globigerinella aequilateralis, CPC30222, CPC30223.

All specimens are from sample 81640021, So16-12SL-075; scale bar 200 μm.

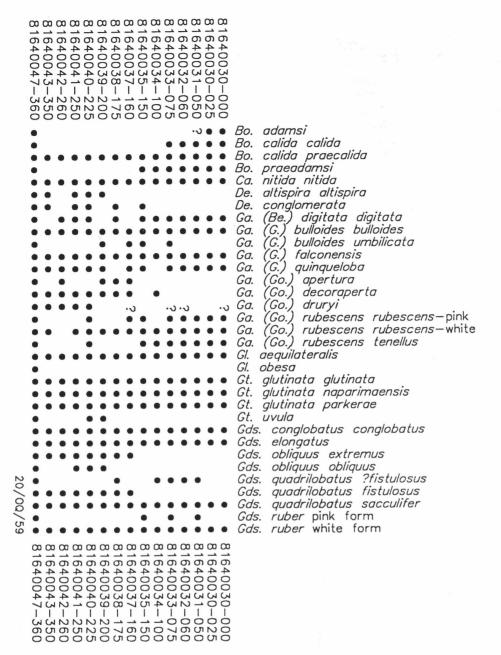
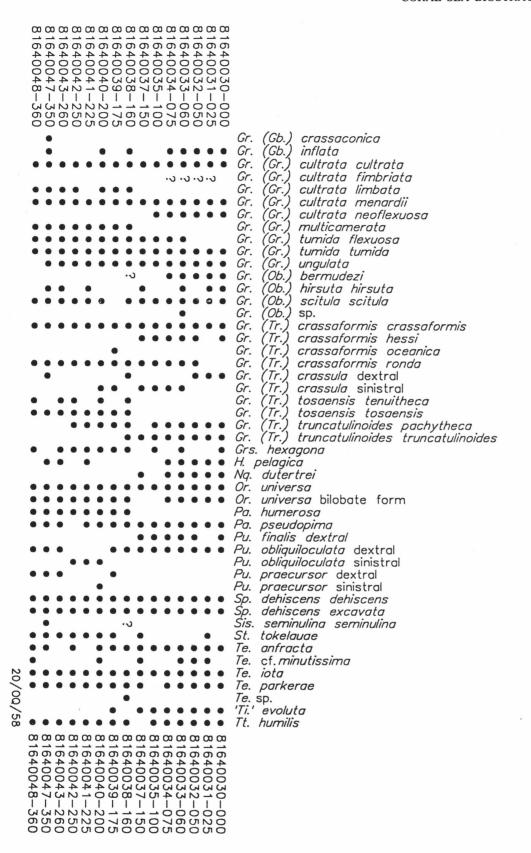


Figure 4. Biostratigraphic range chart for core So16-13SL.

So16-12SL. The lower part of the core (200-242 cm) contains Globorotalia (Truncorotalia) truncatulinoides, Gr. (Tr.) crassaformis hessi, Gr. (Tr.) crassula (sinistral), Gr. (Gr.) tumida flexuosa, Gr. (Gr.) tumida tumida, Gr. (Obandyella) bermudezi, Bolliella praeadamsi n. sp. and Pulleniatina finalis; Gr. (Tr.) tosaensis was not found. At 184 cm Bo. calida calida appears, and the LAD of Gr. (Gr.) tumida flexuosa is at 143 cm. Gr. (Gr.) cultrata fimbriata occurs intermittently from 143 cm to the top of the core. Bolliella adamsi first appears at 74 cm, and at the same time the coiling direction of Gr. (Tr.) crassula changes from sinistral to dextral; populations of Pulleniatina are dextral throughout. Gr. (Globoconella) inflata (including the triangula morphotype) is rare and occurs intermittently through the core. Pink-coloured specimens of Gds. ruber are present throughout the core, but above 242 cm they are rare and sporadic.

So16-13SL. The Pliocene section is from 160 cm to the bottom of the core at 365 cm. The fauna is typified by Globigerinoides

quadrilobatus fistulosus, Gds. obliquus extremus, Globigerina (Globoturborotalita) decoraperta, Globorotalia (Gr.) cultrata limbata, Gr. (Gr.) multicamerata, Gr. (Gr.) tumida flexuosa, Gr. (Truncorotalia) tosaensis and Pulleniatina praecursor. Over the interval 250 to 200 cm, the populations of Pulleniatina change to being dominantly sinistrally coiled; because of the overlap of these sinistral populations with Gr. (Tr.) truncatulinoides, this coiling event almost certainly represents level L5 (1.7 Ma) of Saito (1976). Sphaeroidinellopsis seminulina is present in only one sample at 350 cm. The FAD of Gr. (Tr.) truncatulinoides occurs at 250 cm, and that of both Globigerina (Globoturborotalita) rubescens tenellus and the pink form of Ga. (Go.) rubescens rubescens at 226 cm. The LAD of Dentoglobigerina altispira altispira occurs at 200 cm, coinciding with the highest sinistrally coiled populations of Pulleniatina. At 175 cm 'Tinophodella' evoluta appears. At 160 cm there is an increase in the sand size component in the sediments, accompanied by a faunal change. Populations of Pulleniatina revert to dextral coiling and Globorotalia (Tr.)



crassaformis hessi, Bolliella praeadamsi, Pulleniatina finalis and the pink form of Globigerinoides ruber appear; populations of Gr. (Tr.) crassula are sinistrally coiled. At 74 cm, Bo. calida calida and Gr. (Ob.) bermudezi appear, followed by Bo. adamsi at 50 cm. Above 74 cm, the pink form of Gds. ruber becomes rare and intermittent. Above 50 cm, populations of Gr. (Tr.) crassula are dextrally coiled.

51GC17. The assemblages from the lower part of this core (328–430 cm) contain *Pulleniatina praecursor*, *Gr.* (*Ob.*) bermudezi, *Gr.* (*Tr.*) tosaensis and *Gr.* (*Tr.*) truncatulinoides; populations of *Pulleniatina* are dextrally coiled. The LADs of *Pu. praecursor* and *Gr.* (*Tr.*) tosaensis occur at 328 cm. At 300 cm *Gr.* (*Tr.*) crassaformis hessi appears and populations of *Gr.* (*Tr.*) crassula change from dextral to sinistral coiling. The

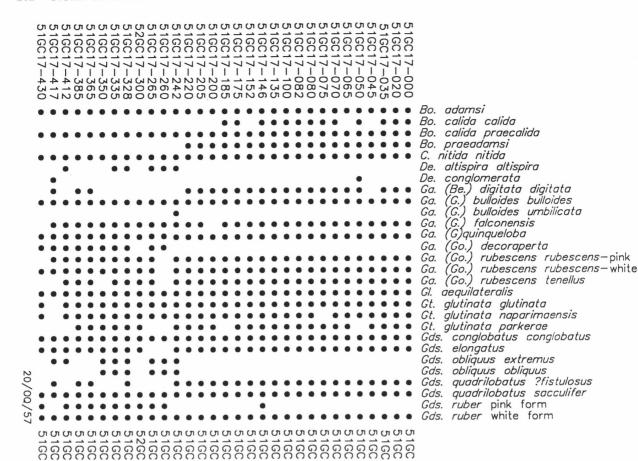


Figure 5. Biostratigraphic range chart for core 51GC17.

-350 -350 -350 -350 -350 -350

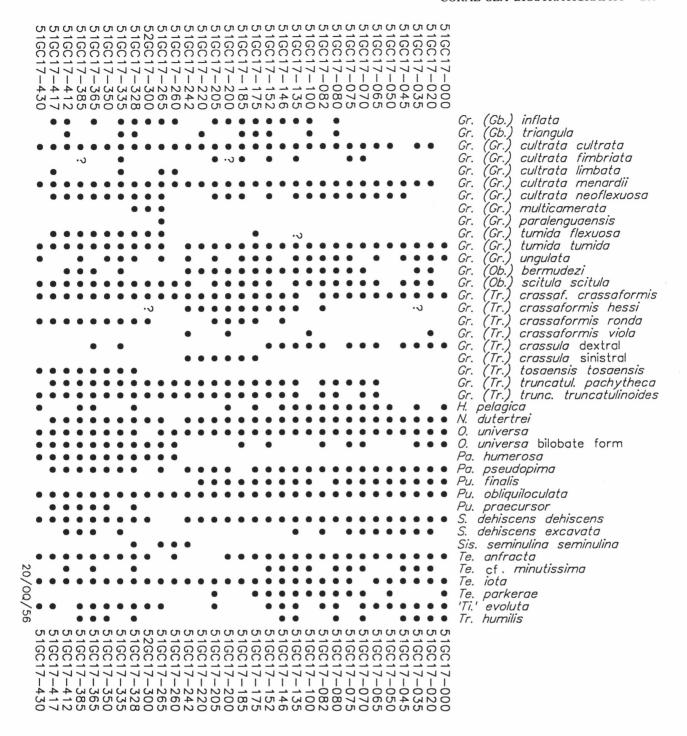
FADs of both Pulleniatina finalis and Bo. praeadamsi are at 220 cm; at 185 cm Bo. calida calida makes its appearance. The LAD of Gr. (Gr.) tumida flexuosa occurs at 175 cm, followed by the LAD of pink-coloured Globigerinoides ruber at 146 cm, although from 260 cm this form becomes very rare. A return to dextrally coiled populations of Gr. (Tr.) crassula takes place at 152 cm. At 82 cm, Bo. adamsi appears. Gr. (Gr.) cultrata fimbriata occurs rarely and intermittently above 335 cm. Between 328 and 242 cm a number of Zone N.17A (late Miocene) species (such as Sphaeroidinellopsis seminulina, Gr. (Gr.) multicamerata, Gr. (Gr.) tumida plesiotumida, Gr. (Gr.) lenguaensis and Globigerina (Go.) nepenthes) are present, indicating reworking. In addition, taxa such as De. altispira altispira, Globigerinoides obliquus extremus, Ga. (Go.) decoraperta and Gr. (Gr.) cultrata limbata are present both below and within this interval, and may also have been reworked from older sediments, but they may be in situ (see below).

51GC18. Pulleniatina finalis, Gr. (Ob.) bermudezi, and Bo. praeadamsi are present throughout the core. The LAD of Gr. (Gr.) tumida flexuosa occurs at 250 cm, and that of pink specimens of Gds. ruber at 180 cm. Bo. calida calida appears at 180 cm, followed by Bo. adamsi at 100 cm. Populations of Gr. (Tr.) crassula are sinistrally coiled in the lower part of the core, changing to dextral coiling above 104 cm. A questionable

occurrence of Gds. obliquus obliquus at 180 cm may be evidence for some reworking.

51GC22. The LADs of *Gr.* (*Tr.*) tosaensis and Pulleniatina praecursor and the FAD of *Gr.* (*Tr.*) crassaformis hessi occur at 214 cm. The FADs of both Bo. praeadamsi and Pu. finalis are at 175 cm, followed by those for Bo. calida calida at 106 cm and Bo. adamsi at 40 cm, the level of the LAD for Gr. (*Tr.*) crassaformis hessi. Pink specimens of Gds. ruber range from the bottom of the core to 50 cm, Gr. (Gr.) tumida flexuosa from the bottom to 80 cm, Gr. (Ob.) bermudezi throughout the core, and Gr. (Gr.) cultrata fimbriata from 125 to 40 cm. Populations of Gr. (Tr.) crassula change from dextral to sinistral coiling at 175 cm, and back to dextral coiling at 70 cm. Globigerina (Go.) decoraperta is restricted to the lowest sample, 218 cm.

51GC45. Bo. praeadamsi and Gr. (Ob.) bermudezi range throughout the core, with Pu. finalis present from 214 cm. Bo. calida calida ranges from 150 cm and Bo. adamsi from 2 cm. Gr. (Gr.) tumida flexuosa, Gr. (Tr.) crassaformis hessi and pink specimens of Gds. ruber range from the bottom of the core, disappearing at 121 cm and 50 cm respectively. Populations of Gr. (Tr.) crassula are dextral from 230 cm, changing to sinistral coiling at 196 cm and reverting to dextral coiling at 2 cm; at two levels within the range of sinistrally coiled



populations, this species is represented by only single dextral specimens. The very short range of *Bo. adamsi* (2 cm) suggests that the uppermost part of the section is missing, perhaps lost during erosion or in coring operations.

76PC06. Gr. (Tr.) crassaformis hessi ranges from the bottom of the core (840 cm) to 230 cm. Both Pu. finalis and Bo. praeadamsi appear at 730 cm, followed by Bo. calida calida at 525 cm and Bo. adamsi at 15 cm. Gr. (Gr.) tumida flexuosa ranges from the base to 190 cm. Sinistrally coiled populations of Gr. (Tr.) crassula range from 485 to 20 cm, but between 720 and 525 cm the species is very rare and intermittent in its distribution. Gr. (Ob.) bermudezi and pink specimens of Gds. ruber are present throughout the core, with the latter suddenly reducing in numbers from approximately 350 cm. The presence

of De. altispira, Globigerinoides obliquus extremus, Gr. (Gr.) cultrata limbata, Gr. (Gr.) multicamerata, Gr. (Gr.) paralenguaensis, Gr. (Tr.) tosaensis, Pu. praecursor and Sis. seminulina indicates reworking throughout the core of latest Miocene and Pliocene faunas. These reworked faunas are confined to turbidites together with magnetic minerals (Peerdeman, ANU, personal communication, 1990). Peerdeman concluded on oxygen isotope evidence that the upper part of the core was lost, and that the top of the core was disturbed, probably during coring operations; a number of turbidites within the core probably coincide with levels where reworked foraminiferids occur. The very short stratigraphic range of Bo. adamsi (which is confined to the top 15 cm) compared with other cores studied, strongly supports the argument for a missing topmost section in this core.

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6666666
 C18-01C
C18-050
C18-059
C18-064
C18-064
C18-089
C18-096
C18-100
C18-110
C18-11
                                                                       1104
                                                                                                                                                                                     Bo. adamsi
                                                                                                                                                                                     Bo. calida calida
                                                                                                                                                                                     Bo. calida praecalida
                                                                                                                                                                                     Bo. praeadamsi
                                                                                                                                                                                      Ca.
                                                                                                                                                                                                     nitida nitida
                                                                                                                                                                                                        conglomerata
                                                                                                                                                                                     De.
                                                                                                                                                                                                        (Be.) digitata digitata
(G.) bulloides bulloides
(G.) falconensis
                                                                                                                                                                                      Ga.
                                                                                                                                                                                      Ga.
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                                                                                                                                                                                                        (G.) quinqueloba
(Go.) rubescens
                                                                                                                                                                                      Ga.
                                                                                                                                                                                                        (Go.) rubescens rubescens pink (Go.) rubescens rubescens white
                                                                                                                                                                                      Ga.
                                                                                                                                                                                      Ga.
                                                                                                                                                                                                        (Go.) rubescens tenellus
                                                                                                                                                                                      Ga
                                                                                                                                                                                      Gl. aequilateralis
                                                                                                                                                                                      Gt.
                                                                                                                                                                                                     glutinata glutinata
                                                                                                                                                                                      Gt.
                                                                                                                                                                                                       glutinata naparimaensis
                                                                                                                                                                                      Gt.
                                                                                                                                                                                                      glutinata parkerae
                                                                                                                                                                                       Gt.
                                                                                                                                                                                                      uvula
                                                                                                                                                                                      Gds. conglobatus conglobatus
                                                                                                                                                                                                          elongatus
                                                                                                                                                                                      Gds.
                                                                                                                                                                                       Gds. obliquus obliquus
                                                                                                                                                                                                          quadrilobatus ?fistulosus
                                                                                                                                                                                      Gds.
                                                                                                                                                                                      Gds.
                                                                                                                                                                                                            quadrilobatus sacculifer
                                                                                                                                                                                       Gds.
                                                                                                                                                                                                          ruber pink form
                                                                                                                                                                                                                                       white form
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                                                                                                                                                                                                            ruber
                                                                                                                                                                                                        (Gb.) inflata
                                                                                                                                                                                      Gr.
                                                                                                                                                                                                      (Gb.) triangula
(Gr.) cultrata
(Gr.) cultrata
                                                                                                                                                                                       Gr.
                                                                                                                                                                                                                             cultrata cultrata
cultrata fimbriata
                                                                                                                                                                                      Gr.
                                                                                                                                                                                      Gr.
                                                                                                                                                                                                      (Gr.)
(Gr.)
(Gr.)
                                                                                                                                                                                       Gr.
                                                                                                                                                                                                                             cultrata menardii
                                                                                                                                                                                       Gr.
                                                                                                                                                                                                                             cultrata neoflexuosa
                                                                                                                                                                                                                             tumida flexuosa
                                                                                                                                                                                      Gr.
                                                                                                                                                                                                       (Gr.) tumida tumida
(Gr.) ungulata
                                                                                                                                                                                      Gr.
                                                                                                                                                                                      Gr.
                                                                                                                                                                                      Gr.
Gr.
                                                                                                                                                                                                       (Ob.) bermudezi
(Ob.) scitula scitula
                                                                                                                                                                                                       (Tr.
                                                                                                                                                                                      Gr.
                                                                                                                                                                                                                             crassaformis crassaformis
                                                                                                                                                                                                       (Tr.) crassaformis hessi
                                                                                                                                                                                      Gr.
                                                                                                                                                                                                      (Tr.) crassaformis ronda
(Tr.) crassula dextral
                                                                                                                                                                                      Gr.
                                                                                                                                                                                      Gr.
                                                                                                                                                                                                      (Tr.) crassula sinistral
                                                                                                                                                                                      Gr.
                                                                                                                                                                                     Gr.
                                                                                                                                                                                                     (Tr.)
(Tr.)
                                                                                                                                                                                                                            truncatulinoides pachytheca
truncatulinoides truncatulinoides
                                                                                                                                                                                      Gr.
                                                                                                                                                                                     H. pelagica
                                                                                                                                                                                     Nq.
                                                                                                                                                                                                    dutertrei
                                                                                                                                                                                     Or.
                                                                                                                                                                                                      universa
                                                                                                                                                                                     Or.
                                                                                                                                                                                                     universa bilobate form
                                                                                                                                                                                     Pa.
                                                                                                                                                                                                      humerosa
                                                                                                                                                                                     Pa. pseudopima
                                                                                                                                                                                     Pu. finalis
                                                                                                                                                                                     Pu.
                                                                                                                                                                                                      obliquiloculata
                                                                                                                                                                                     Sp.
Sp.
St.
                                                                                                                                                                                                      dehiscens dehiscens
                                                                                                                                                                                                      dehiscens excavata
                                                                                                                                                                                                      tokelauae
                                                                                                                                                                                      Te.
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                                                                                                                                                                                                     parkerae
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                                                                                                                                                                                      'Ti.
                                                                                                                                                                                                      evoluta
                                                                                                                                                                                      Tu. humilis
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Figure 6. Biostratigraphic range chart for core 51GC18.

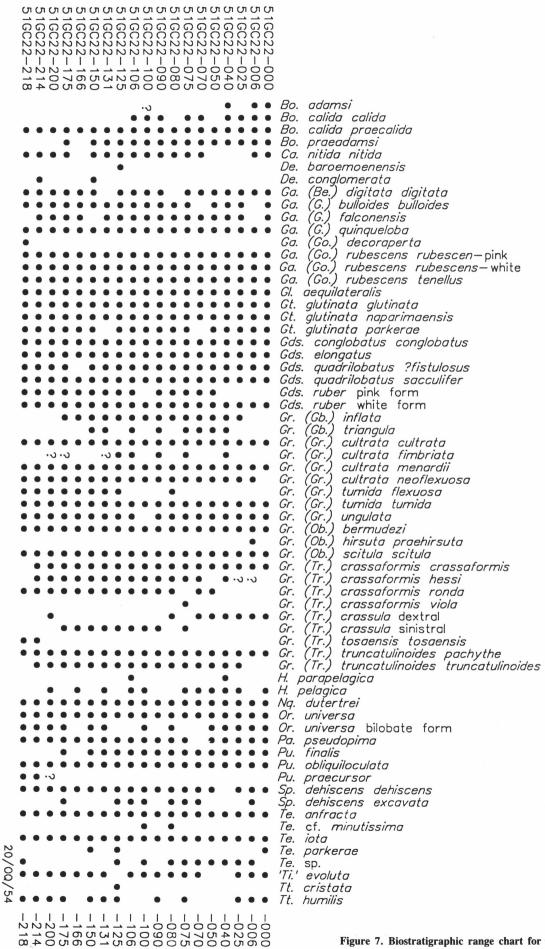


Figure 7. Biostratigraphic range chart for core 51GC22.

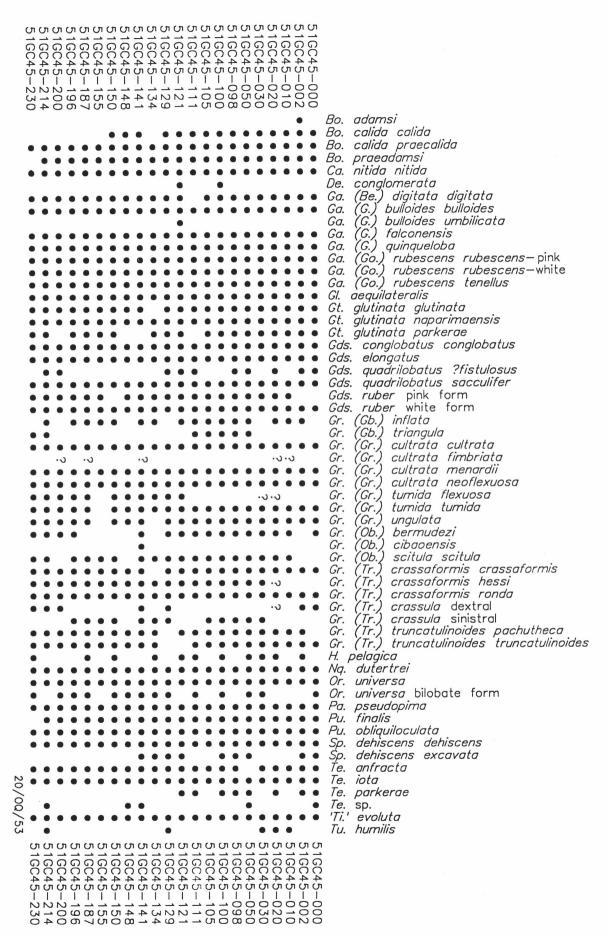


Figure 8. Biostratigraphic range chart for core 51GC45.

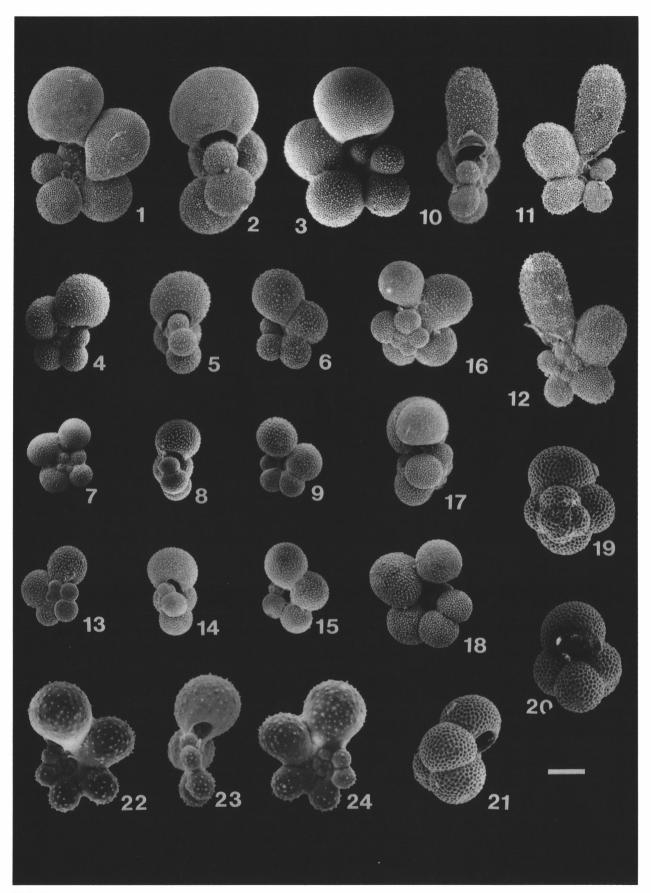


Plate 2.
1–9, Bolliella praeadamsi n. sp., paratypes, CPC30224, CPC30225, CPC30226. 10–12, Bolliella adamsi, CPC30227. 13–15, Bolliella calida praecalida, CPC30228. 16–18, Bolliella calida calida, CPC30229. 19–21, Globigerina (Globoturborotalita) decoraperta, CPC30230. 22–24, 'Tinophodella' evoluta, CPC30231.
Figs 1–18, sample 81640021, So16-12SL-075, scale bar 180 μm; 19–21, sample 51GC17-260, scale bar 100 μm; 22–24, sample 51GC45-100, scale bar 60 μm.

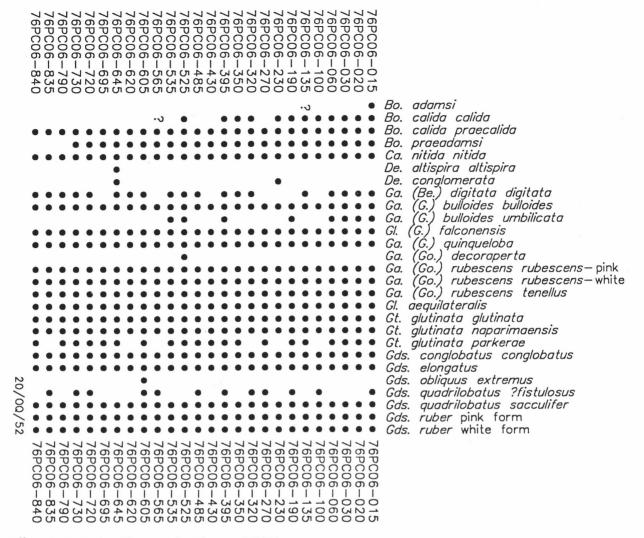


Figure 9. Biostratigraphic range chart for core 76PC06.

Pleistocene biostratigraphy

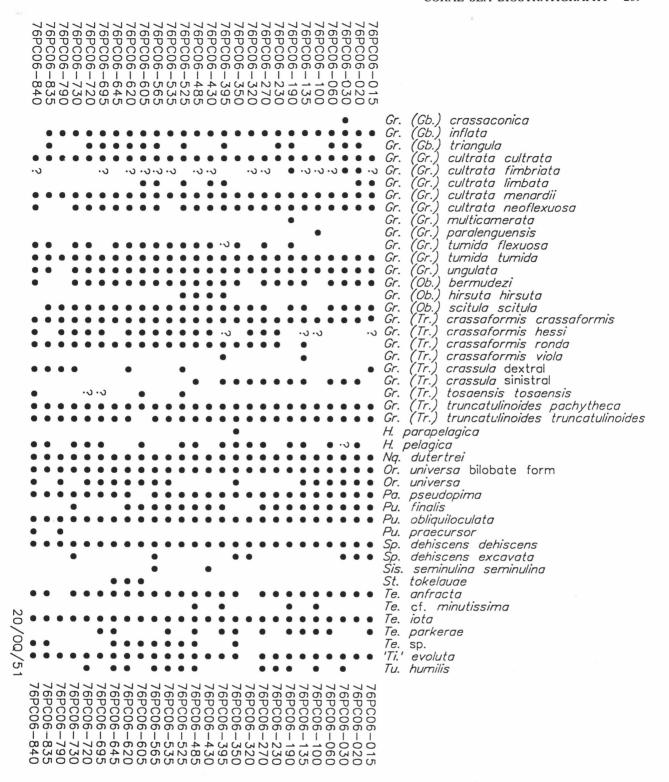
Examination of the literature indicates that there are 23 potential planktic foraminiferal events within the Pleistocene for the Indo-Pacific region (Table 1); in addition, four others have been revealed by this study (Table 1). Those used herein are given in Table 2. Of the 27 events, 9 have been correlated to the oxygen isotopic stratigraphic sequence in core 76PC06 (Fig. 10). The remaining events are from the sequence below 76PC06 and so cannot be correlated with the oxygen isotope sequence. In addition, some of the events listed in Table 1 are from levels within the early Pleistocene hiatus and so could not be evaluated in this study. As discussed below, one of these, the FAD of Pulleniatina finalis, has been given ages which differ from those proposed by some previous workers. Though some of these events have been used in other areas (e.g. the FADs of Bo. calida calida and Gr. (Tr.) crassaformis hessi), a number have not (e.g. the FADs of Bo. praeadamsi and Bo. adamsi, and the coiling change in Gr. (Tr.) crassula).

The composition of planktic foraminiferal assemblages in the cores from northeastern Australia is constant, a feature to be expected if the region has remained in the tropical-subtropical belt during the Pleistocene. Thus zones of influx of cooler forms are absent from the cores; cool water species (such as Gr. (Globoconella) inflata, Gr. (Gb.) triangula and Globigerina (Globigerina) bulloides) are present but are al-

Table 1. Planktic foraminiferal biostratigraphic events of potential use for Pleistocene biostratigraphy.

```
LAD Globorotalia (Truncorotalia) crassaformis hessi
FAD Bolliella adamsi
Change from s to d coiling in Gr. (Tr.) crassula
LAD Gr. (Gr.) tumida flexuosa
LAD Globigerinoides ruber pink form
FAD Bolliella calida calida
LAD Globigerina (Globoturborotalita) decoraperta
FAD Bolliella praeadamsi
FAD Pulleniatina finalis
Change from d to s coiling in Gr. (Tr.) crassula
LAD Gr. (Tr.) tosaensis
LAD Pu. praecursor
FAD Gr. (Tr.) crassaformis hessi
FAD Gr. (Ob.) bermudezi
L1 coiling interval for Pulleniatina
L2 coiling interval for Pulleniatina
L3 coiling interval for Pulleniatina
LAD Gds. quadrilobatus fistulosus
LA coiling interval for Pulleniatina
LAD Gds. obliquus extremus
FAD 'Tinophodella' evoluta
LAD Gds. obliquus obliquus
LAD Dentoglobigerina altispira altispira
FAD Ga. (Globoturborotalita) rubescens pink form L5 coiling interval for Pulleniatina
FAD Ga. (Go.) rubescens tenellus
FAD Gr. (Tr.) truncatulinoides
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ways rare and form a very small component in the assemblages. This makes it impossible to utilise the presence or

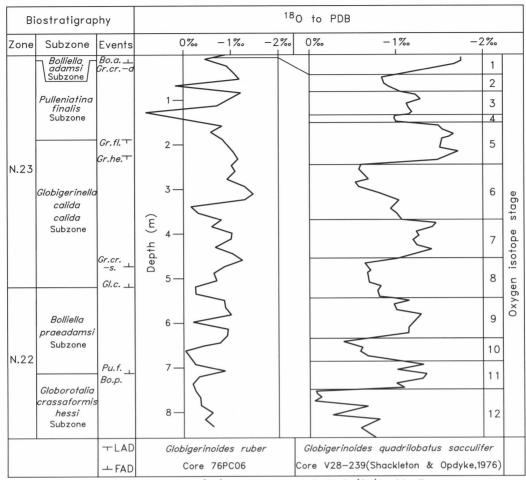


absence of temperature-controlled forms in the Coral Sea area as has been possible in other places (such as the Gulf of Mexico).

Reworking is obvious in some cores, particularly in 51GC17 and 76PC06. In most cases this represents reworking from the late Pliocene (forms such as *Gr. (Tr.) tosaensis* and *Pu. praecursor*) although some from the late Miocene (*Gr. (Gr.) paralenguaensis* and *Gr. (Gr.) tumida plesiotumida*) is evident. Reworking does not pose problems for the biostratigraphy of the middle and late Pleistocene, as all species involved had become extinct by the lowermost Pleistocene.

Evolutionary bioseries have provided useful markers for this study, in particular the *Bo. calida praecalida–Bo. calida calida*, the *Bo. calida praecalida–Bo. praeadamsi–Bo. adamsi* and *Gr. (Tr.) crassaformis ronda–Gr. (Tr.) crassaformis hessi* lineages. A change in coiling dominance in populations of *Gr. (Tr.) crassula* has also been particularly useful.

All of the cores studied contain the section above the FAD of *Bo. praeadamsi*. In this part, the events shown in Table 1 consistently occur in the same order, testifying to their reliability, at least for the Coral Sea region adjacent to northern Australia. Two cores (So16-13SL and 76PC06) penetrated into



Bo.a.-Bolliella adamsi Gr.cr.-d.-Gr.(Tr.)crassula-dextral Gr.fl.-Gr.(Gr.)tumida flexuosa Gr.he.-Gr.(Tr.)crassaformis hessi Gr.cr.-s-Gr.Tr.)crassula-sinistral Gl.c.-Globigerinella calida calida Pu.f.-Pulleniatina finalis Bo.p.-Bolliella praedamsi

Figure 10. Relationship of biostratigraphic events and oxygen isotope stratigraphy in core 76PC06. Oxygen isotope data from Peerdeman (ANU, personal communication).

that part of the section above the FAD of Gr. (Tr.) crassaformis hessi, and two others (51GC17 and 51GC22) occur below that event. In So16-13SL the latest Pliocene disconformably underlies the Gr. (Tr.) crassaformis hessi Subzone. Again, all events recognised in this area occurred in the same order.

Biostratigraphic events

LAD of Globorotalia (Truncorotalia) crassaformis hessi. This event takes place with the top of Zone N.22, above the FAD of Bo. calida calida and below the FADs of both Bo adamsi and Gr. (Gr.) tumida flexuosa (Bolli & Saunders, 1985). In this study, the LAD of Gr. (Tr.) crassaformis hessi occurs above these events in all sections studied except 76PC06, where it occurs within the range of sinistrally coiled Gr. (Tr.) crassula, between the LAD of Gr. (Gr.) tumida flexuosa and the FAD of Bo. adamsi, within oxygen isotope stage 5. However, the top part of this core was lost during coring operations which also disturbed the topmost section. On the evidence from other cores, this FAD event is probably within oxygen isotope stage 2.

FAD Bolliella adamsi. This event occurs within the later Quaternary (Blow, 1969; Saito & others, 1981; Bolli & Saunders, 1985). Bolli & Saunders (1985, fig. 11) indicate that Bo. adamsi is restricted to the Holocene. This species is found

Table 2. Biostratigraphic events from the middle Gauss Chron to present used in this study.

1. LAD Globorotalia (Truncorotalia) crassaformis	1. 1	. LAI	AD Globorotalia	(Truncorotalia)	crassaformis	hessi
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2. FAD Bolliella adamsi

Change from s to d coiling in Gr. (Tr.) crassula LAD Gr. (Gr.) tumida flexuosa

LAD Gds. ruber pink form

FAD Bo. calida calida

LAD Ga. (Go.) decoraperta

FAD Bo. praeadamsi 9. FAD Pu. finalis

10. Change from d to s coiling in Gr. (Tr.) crassula

11. LAD Gr. (Tr.) tosaensis 12. LAD Pu. praecursor

13. FAD Gr. (Tr.) crassaformis hessi

14. FAD Gr. (Ob.) bermudezi

Zonal markers

Zone N19/20-N.21 boundary: FAD Gr. (Tr.) tosaensis Zone N.21-N.22 boundary: FAD Gr. (Tr.) truncatulinoides

Zone N.22-N.23 boundary: FAD Bo. calida calida

Subzonal markers

Zone N.22 Subzones

Gds. quadrilobatus fistulosus subzone: FAD Gr. (Tr.) truncatulinoides Gr. (Tr.) crassaformis viola subzone: LAD Gds. quadrilobatus fistulosus Gr. (Tr.) crassaformis hessi subzone: FAD Gr. (Tr.) crassaformis hessi Bo. praeadamsi subzone: FAD Bo. praeadamsi

Zone N.23 Subzones

Bo. calida subzone: FAD Bo. calida calida Pu. finalis subzone: LAD Gr (Gr.) tumida flexuosa

Bo. adamsi subzone: FAD Bo. adamsi

only in the upper parts of the cores studied from northeastern Australia, above the FAD of Bo. calida calida, and above the interval of sinistrally coiled Gr. (Tr.) crassula. The species occurs only in the top 15 cm of 76PC06, which is within either oxygen isotope stage 1 or 2 in the Holocene. I have used this event to define the base of the Bolliella adamsi Subzone (Chaproniere, 1985b).

Coiling changes in populations of Globorotalia (Truncorotalia) crassula. A change in coiling direction in populations of Gr. (Tr.) crassula from dominantly dextral to sinistral and then the reverse has been noted for most sections, even though the absence or rarity of this species at some levels make it difficult to accurately locate the level where the changes occur. Although populations of this species appear to be dominated by dextrally coiled individuals through most of its range, at a level immediately before the FADs of Bolliella praeadamsi and Pulleniatina finalis there is a rapid shift from dextral to sinistral coiling. Sinistrally coiled populations continue until a level between the LAD of Gr. (Gr.) tumida flexuosa and the FAD of Bo. adamsi, when the populations revert to dextral coiling. In core 76PC06 this species occurs intermittently in the lower part of the section. Though the change from dextral to sinistral coiling occurs immediately above the FAD of Bo. calida calida (in oxygen isotope stage 7), the specimens are extremely rare and intermittent. The apparent coiling change is therefore unreliable, especially when compared with data from other cores. In cores So16-13SL, 51GC22 and 51GC45 the FAD of Gr. (Tr.) crassula occurs above the base of the Bo. praeadamsi Subzone, and in cores So16-12SL and 51GC18 sinistral Gr. (Tr.) crassula ranges from within this subzone; in core 51GC17 this event occurs within the top of the Gr. (Tr.) crassaformis hessi Subzone. Thus, the evidence suggests that the change from dextral to sinistral coiling takes place near the boundary of the Gr. (Tr.) crassaformis hessi and Bo. praeadamsi Subzones, below oxygen isotope stage 10, possibly within oxygen isotope stage 11. The change from sinistral to dextral coiling takes place between the LAD of Gr. (Gr.) tumida flexuosa and the FAD of Bo. adamsi; this event occurs within a turbidite interval near the top of core 76PC06, making the correlation with the oxygen isotope stage chronology unreliable (Peerdeman, ANU, personal communication, 1990). Notwithstanding, this event must take place within the interval of isotope stages 1 to 4 in this core, probably within oxygen isotope stages 2 or 3. This interval of sinistral coiling in the later Pleistocene has not been recognised before.

LAD Globorotalia (Globorotalia) tumida flexuosa. Blow (1969) believed that this form did not range into Zone N.23 and so did not overlap with Bo. calida calida. However, Bolli & Premoli Silva (1973) used the overlap of the two taxa to define their Globigerina calida calida Subzone in the Caribbean region. Bolli & Saunders (1985) stated that Gr. (Gr.) tumida flexuosa had a very limited stratigraphic range in the Indo-Pacific region where it was confined to the early Pliocene. However, it has the same range in the vicinity of Wallis Island as that recorded from the Caribbean region (Chaproniere, 1985b; present study). The position of this event is contrary to that recorded by Kennett & Srinivasan (1983), who locate it at the Zone N.21-N.22 boundary. The highest occurrence of this subspecies is at 190 cm in core 76PC06, within oxygen isotope stage 4.

LAD Globigerinoides ruber pink form. Thompson & others (1979) concluded that this event occurred at 120 000 years BP in the Indian and Pacific Oceans, even though it is extant in the Atlantic Ocean. It could therefore be valuable for biostratigraphy for the region. I have recorded rare specimens in Holocene sediments both in the present study and near Wallis Island, north of Fiji (Chaproniere, 1985b) and the Tongan region (Chaproniere, 1985a; in press). The specimens suggest either that it is extant in the region, or that it has been reworked from older sediments over wide areas of the subtropical-tropical southwestern Pacific. The rarity and patchy distribution of this readily identified form favour reworking. In most cores there is a rapid reduction of population size of pink individuals at levels above the FAD of Bo. calida calida and below the LAD of Gr. (Gr.) tumida flexuosa, apparently very close to the 120 000 years BP date (which corresponds to within oxygen isotope stage 5e) obtained by Thompson & others (1979). The reduction in numbers (top acme) appears to provide a useful event for the region. In core 76PC06 the sudden reduction in numbers of pink Gds. ruber occurs at 350 cm, within the lower part of oxygen isotope stage 5.

FAD Bolliella calida calida. Blow (1969) used this event to define the base of Zone N.23 which he considered to range from late Pleistocene to the Holocene. Bolli & Premoli Silva (1973) believed that the FAD of this subspecies occurred at 200 000 yrs. BP (within oxygen isotope stage 7), that is, within the latest Pleistocene. This is a similar level to that recorded in 76PC06, where the event occurs in oxygen isotope stage 7. I have strictly adhered to Blow's (1969) criteria for the separation between calida and praecalida morphotypes; thus, calida has a test with at least 4.5 chambers in the final whorl, and the chambers show distinct radial elongation.

LAD Globigerina (Globoturborotalita) decoraperta. This event coincides with the Zone N.21-N.22 boundary (Kennett & Srinivasan, 1983). The species is rare in the cores studied, but it persists to higher levels than previously recorded. Of the three cores from which it has been recorded, only in core So16-13SL (where it ranges into the Bo. praeadamsi Subzone) can reworking from older levels be rejected. In core 76PC06 it ranges into the Bo. calida calida Subzone, but is associated with reworked taxa. In core 51GC17 it ranges into the Gr. crassaformis hessi Subzone. It is sometimes but not always accompanied with reworked forms, which suggests that it is probably in situ in this section.

FAD Bolliella praeadamsi. This event consistently occurs at the same time as the FAD of Pulleniatina finalis, below oxygen isotope stage 10, and probably within isotopic stage 11 in core 76PC06.

FAD Pulleniatina finalis. There is some disagreement in the literature as to the biostratigraphic range of this species. Banner & Blow (1967) described this form from the Recent of the Atlantic Ocean, but Blow (1969) considered that it ranged from well within the Pleistocene to the present day. Subsequently, Saito & others (1975) believed its FAD to be within the lower part of the Pleistocene. Poag & Valentine (1976) found it in Subzone B of their Turborotalia inflata IV Bioecozone, which they correlated to a level within the Jaramillo Subchron, above the level given by Saito & others (1975). Lamb & Beard (1972) believed this event occurred within the Illinoian Glacial Stage, which ranges from oxygen isotope stages 16 to 13 (Thunell, 1984, table 2.2), and correlates with levels in the lower part of the Brunhes Chron. Bolli & Saunders (1985) claim that it appeared within the Pleistocene in the Atlantic Province, and much earlier in the Indo-Pacific Province based on evidence credited to Beckmann (1971) from the central Pacific. However, Beckmann (in Tracey & others, 1971a, p. 292; 1971b, p. 466) shows that the species is restricted to the Pleistocene, and gives no evidence to support the contention made by Bolli & Saunders (1985). This record of an earlier FAD is therefore discounted here.

Bolli (1966) and Belford (1988) noted that forms close to Pu. finalis occur in the Pliocene. These records were attributed to

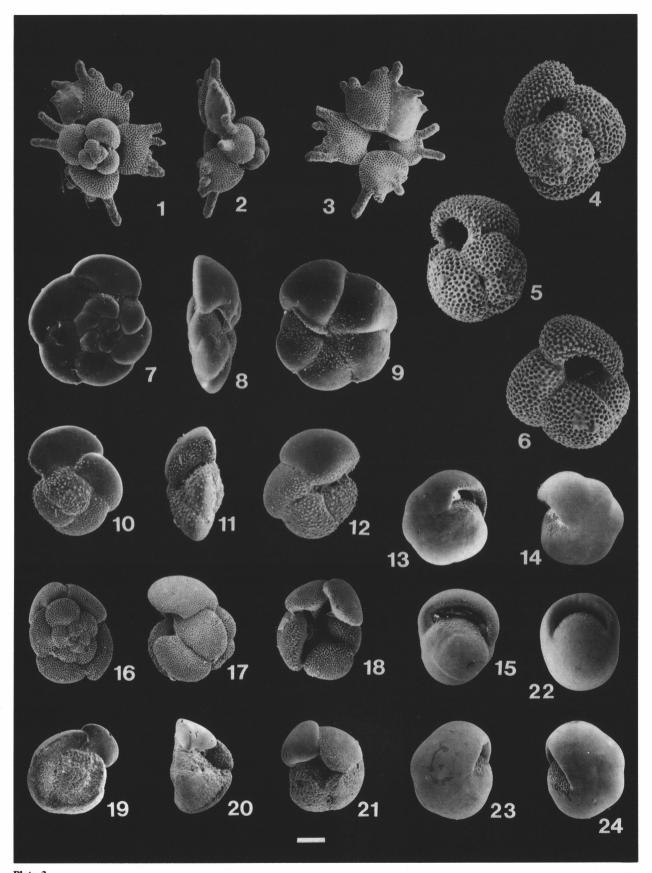


Plate 3.

1-3, Globigerinoides quadrilobatus fistulosus, CPC30232, sample 81640043, So16-13SL-260. 4-6, Globigerinoides obliquus extremus, CPC30233, sample 81640030, So16-13SL-000. 7-9, Globorotalia (Obandyella) bermudezi, CPC30234, sample 51GC45-105. 10-12, Globorotalia (Truncorotalia) crassula, CPC30235, sample 81640034, So16-13SL-075. 13-15, Pulleniatina praecursor, CPC30236, sample 51GC22-214. 16-18, Dentoglobigerina altispira altispira, CPC30237, sample 81640040, So16-13SL-200. 19-21, Globorotalia (Truncorotalia) crassaformis hessi, CPC30238, sample 51GC18-050. 22-24, Pulleniatina finalis, CPC30239, sample 76PC06-730.

Scale bar figs 1-3, 250 μm; 4-9, 75 μm; 10-12, 135 μm; 13-24, 200 μm.

environmental factors by Bolli & Saunders (1985), but the presence of these Pu. finalis-like forms in the early Pliocene suggests that they represent a short-lived side branch from Pu. obliquiloculata which was not related to the later Pu. finalisstage descendant. Thus, though there is good evidence for the FAD of Pu. finalis within the Pleistocene, there is no general consensus on the exact level at which Pu. finalis first appears. This lack of consensus is probably the result of differing taxonomic interpretations of this species due to difficulties in interpreting the degree of involution of the test. In the sections used in this study, Pu. finalis first appears at a level approximating oxygen isotope stage 12, which is close to that recorded by Lamb & Beard (1972) from the Caribbean region, suggesting that the contention of Bolli & Saunders (1985) is incorrect. As noted above, Saito & others (1975) believed that the FAD of Pu. finalis occurred earlier in the Pleistocene than that recorded by Lamb & Beard (1972), Poag & Valentine (1976), or in this study. Saito & others (1975) failed to illustrate their concept of the taxon, and the illustration given by Saito & others (1981) is of specimens which appear to have some of the earlier spire visible, and are probably not entirely involute; in addition they are from the Holocene. Because Banner & Blow (1967) described finalis as being involute, the specimen illustrated by Saito & others (1981) should not be referred to this species, but to Pu. obliquiloculata. The specimens of Pu. finalis illustrated by Poag & Valentine (1976) fall within my interpretation of the species, but they are from the Holocene rather than from the level of its FAD in the Louisiana shelf area. The level of FAD of involute morphotypes of Pulleniatina is still uncertain, but it seems that the records of Saito & others (1975) and Bolli & Saunders (1985) are not reliable.

LAD of Globorotalia (Truncorotalia) tosaensis. This event was used by Kennett and Srinivasan (1983) to mark the top of their Gr. truncatulinoides-Gr. tosaensis overlap zone. As noted above, this event appears to coincide with the FAD of Gr. (Tr.) crassaformis hessi. The event occurs in oxygen isotope stage 15 (Thompson & Sciarrillo, 1978).

LAD Pulleniatina praecursor. Pu. praecursor has been found in four cores. In So 16-13SL it occurs within the lower part of the core, where it overlaps with Gr. (Tr.) truncatulinoides and so clearly ranges within the lower part of Zone N.22, higher than the Zone N.21 level indicated by Kennett & Srinivasan (1983). In core 51GC17 it ranges throughout the Gr. (Tr.) crassaformis viola Subzone, in 51GC22 into the basal part of the Gr. (Tr.) crassaformis hessi Subzone and in 76PC06 into the upper part of the Gr. (Tr.) crassaformis hessi Subzone. In core 51GC17 reworked late Miocene faunas are present at the same levels, and in core 76PC06 reworked late Pliocene to early Pleistocene faunas are present, suggesting that the specimens of Pu. praecursor may also be reworked. Because the FAD of Pu. praecursor occurs within Zone N.19/20 (Kennett & Srinivasan, 1983) it is unlikely that its presence in core 51GC17 is due to reworking, as the reworked species are from Zone N.17. However, in core 76PC06 this species occurs with taxa derived from levels above Zone N.19/20, above its FAD, suggesting that in this section it may have been reworked. In core 51GC22, however, Pu. praecursor is present without reworked older faunas, suggesting that it is in situ. Thus the evidence suggests that this species ranges at least into the early Pleistocene, but may also range as high as the base of the Gr. (Tr.) crassaformis hessi Subzone, that is, into the basal Brunhes Chron.

FAD of Globorotalia (Truncorotalia) crassaformis hessi. This event was used by Bolli & Premoli Silva (1973) to define the base of their Gr. crassaformis hessi Subzone. In the studied sections this event always pre-dates the FAD of Bo. calida

calida and post-dates the FAD of Gr. (Tr.) truncatulinoides. Bolli & Saunders (1985, fig. 11) show that the LAD of Gr. (Tr.) tosaensis pre-dates the FAD of Gr. (Tr.) crassaformis hessi, but in the three sections which include both events (51GC17, 51GC22, 76PC06) there is a brief overlap between the two taxa in one (51GC22) and in another (51GC17) no such overlap is observed. In the third there is considerable overlap between the two, but a number of latest Miocene or Pliocene forms (such as De. altispira altispira, Gds. obliquus extremus, Gr. (Gr.) multicamerata, Gr. (Gr.) paralenguaensis, and Sis. seminulina) are also present, indicating reworking. The presence of Gr. (Tr.) tosaensis at these levels is therefore questionable. Thus, this work suggests that both the events take place at approximately the same time, but with little overlap. For this reason I have provisionally placed this event within oxygen isotope stage 17.

FAD Globorotalia (Obandyella) bermudezi. Kennett & Srinivasan (1983) and Rögl & Bolli (1973) placed this event at the Zone N.21-N.22 boundary. The species is common in most samples from the cores studied, although its distribution is discontinuous in the lower part of its range. In some cores (51GC17, 51GC22) this event occurs within the top of the Gr. crassaformis viola Subzone. In others, which penetrate into only the younger part of the section, Gr. (Ob.) bermudezi ranges from the bottom. Although this event appears to occur within the Gr. (Tr.) crassaformis viola Subzone near the base of the Brunhes Chron, the absence of samples from that part of the Pleistocene below the Brunhes Chron makes it necessary to await other studies from levels lower in the Pleistocene before a full evaluation can be made.

Coiling changes in populations of Pulleniatina. Saito (1976) recognised five intervals where populations of Pulleniatina were dominated by sinistrally coiled individuals within Zone N.22. Though these intervals were short they could be correlated globally, based on palaeomagnetic data (Fig. 2). The sole interval of sinistrally coiled populations encountered in this study is in core So16-13SL, where they occur over a 50 cm interval coinciding with the overlap of Gds. quadrilobatus fistulosus and Gr. (Tr.) truncatulinoides near the LAD of the latter. This is at the same position as that recorded by Saito (1976) as the L5 coiling interval of Pulleniatina; this coincides with the base of the Olduvai Subchron, and is correlated to that interval here.

LAD Globigerinoides quadrilobatus fistulosus. The FAD of Gr. (Tr.) truncatulinoides and the LAD of Gds. quadrilobatus fistulosus coincide with the Zone N.21-N.22 boundary (Kennett & Srinivasan, 1983) even though the FAD of Gr. (Tr.) truncatulinoides occurs just below the base of the Olduvai event, whilst the LAD of Gds. quadrilobatus fistulosus occurs just above the top of the Olduvai event (Berggren & others, 1980). In this work, the LAD of Gds. quadrilobatus fistulosus occurs well within the lower part of Zone N.22. This is similar to the LAD recorded by Berggren & others (1985), who indicated that this event occurs immediately above the Olduvai Subchron at 1.6 Ma. In core So16-13SL this event coincides with a hiatus.

LAD Globigerinoides obliquus extremus. Kennett & Srinivasan (1983) equated this event with the Zone N.21-N.22 boundary, but Berggren & others (1980, 1985) placed it within the lower part of the Olduvai Subchron, within Zone N.22. In core So16-13SL, this subspecies is found to overlap with Gr. (Tr.) truncatulinoides and Gds. quadrilobatus fistulosus, supporting the view of Berggren & others (1980, 1985). This taxon occurs at even higher levels in two other cores, 76PC06 (in the Bo. praeadamsi Subzone) and 51GC17 (where it is present in the Gr. (Tr.) crassaformis hessi Subzone), but in both instances reworking can be demonstrated.

FAD 'Tinophodella' evoluta. This minute form was described from DSDP Site 208 by Fordham (1986) who considered that it was probably restricted to the Pleistocene. This species occurs in the fine fractions (<250 μm) of all cores studied. In core So16-13SL this species was found to occur from 175 cm and above, ranging from a level within the Gds. quadrilobatus fistulosus Subzone. The position of this event requires assessment in other areas to test its full biostratigraphic potential.

LAD Globigerinoides obliquus obliquus. This event occurs within the top part of Zone N.21, below the FAD of Gr. (Tr.) truncatulinoides, and below the LAD of Gds. obliquus extremus (Kennett & Srinivasan, 1983). In core So16-13SL, the LAD of Gds. obliquus obliquus occurs above the FAD of Gr. (Tr.) truncatulinoides, below the LADs of Gds. quadrilobatus fistulosus and Gds. obliquus extremus, and within the range of sinistrally coiled populations of Pulleniatina. These data indicate that this event takes place within the Olduvai Subchron in the Coral Sea region. Specimens of Gds. obliquus obliquus are also present in core 51GC17 at levels containing a number of reworked late Miocene taxa; these specimens are also believed to have been reworked.

LAD Dentoglobigerina altispira altispira. This event took place within the lower part of Zone N.21 (Kennett & Srinivasan, 1983), and is dated at 2.9 Ma (Berggren & others, 1985), the same level as the FAD of Gds. quadrilobatus fistulosus. Within core So16-13SL, this subspecies occurs with Gr. (Tr.) truncatulinoides, Gds. quadrilobatus fistulosus and Gds. obliquus extremus, indicating that it ranges to within the basal part of Zone N.22. There is no evidence for reworking in this core, although the presence of De. altispira altispira in core 51GC17 is ascribed to reworking. It is absent from the Pliocene in cores 51GC73 and 51GC74.

FAD Globigerina (Globoturborotalita) rubescens pink form. Pink forms of Ga. (Go.) rubescens seem to develop within the Pleistocene (Parker, 1967; Jenkins & Orr, 1972). However, the reliability of the FAD of the pink coloured form has not yet been fully established. I have recorded it as ranging from within the Gr. (Tr.) crassaformis viola Subzone near Wallis Island (Chaproniere, 1985b), and a similar range is found in the cores used in this study. In core So16-13SL, the FAD of pink forms occurs 20 cm above the FAD of Gr. (Tr.) truncatulinoides, within the range of Gds. quadrilobatus fistulosus and sinistrally coiled populations of Pulleniatina. As noted above, this assemblage correlates with the lower part of the Olduvai Subchron according to Saito (1976), suggesting that the FAD of pink forms of Ga. (Go.) rubescens occurs at this level, within the basal part of Zone N.22, in the latest Pliocene. Even though this event appears to have potential for marking the base of Zone N.22, it requires verification at additional locations.

FAD Globigerina (Globoturborotalita) rubescens tenellus. Kennett & Srinivasan (1983) place this event within the top of Zone N.21 (late Pliocene), but Rio & others (1984a) show it occurring above the base of Zone N.22. In core So16-13SL this event occurs 24 cm above the FAD of Gr. (Tr.) truncatulinoides, within the range of Gds. quadrilobatus fistulosus and sinistrally coiled populations of Pulleniatina, indicating that the FAD of Ga. (Go.) rubescens tenellus occurs within the lower part of the Olduvai Subchron and the lower part of Zone N.22, similar to that recorded by Rio & others (1984a).

FAD of *Globorotalia* (*Truncorotalia*) *truncatulinoides*. As noted above, it is uncertain whether this event took place at the same time in lower and higher latitudes, although it seems constant globally for low latitudes. The event has been dated as

1.9 Ma and approximates the base of the Olduvai magnetic event (Berggren & others, 1985).

Discussion

Of the 27 events discussed above (Table 1), 23 have been recognised by earlier workers, and only four are newly recognised. Few of these have been used biostratigraphically, at least in the Indo-Pacific region, where there have been few attempts at a Pleistocene planktic foraminiferal biostratigraphy. Many of the events have been found at different levels to those previously recorded, which may be due in part to the paucity of studies in subtropical-tropical sequences, and in part to differences in taxonomic usage. Some of these events occur either in the 'missing' part of Pleistocene represented by a hiatus encountered in core So16-13SL, or at a level below that penetrated by most of the cores. Nine of the events (1 to 9 in Table 2) are present in core 76PC06, the only core for which oxygen isotope data are available (Fig. 10; Peerdeman, ANU, personal communication, 1990), and have been correlated with the oxygen isotope stratigraphic scheme. Because of the lack of information on many of these events within the western Pacific region I have used for subzonal definitions only those events which have been found elsewhere and appear to be chronologically consistent.

Biostratigraphic zonation

As noted above, the zonal scheme of Blow (1969) forms the basis for the zonation used for this study. This scheme uses the events of the FADs of *Gr.* (*Tr.*) tosaensis, *Gr.* (*Tr.*) truncatulinoides and *Bo.* calida calida to define the bases of Zones N.21, N.22 and N.23 respectively. Bolli & Premoli Silva (1973) subdivided the last two zones into five Subzones. I found this not to be fully applicable to the region adjacent to Wallis Island (Chaproniere, 1985b), and a modified subzonal scheme was proposed. However this also proved inadequate for the area covered by this study, and so has once again been modified (Fig. 11).

Zone N.21. The lower section of core So16-13SL contains Gr. (Tr.) tosaensis and Gds. quadrilobatus fistulosus, without Gr. (Tr.) truncatulinoides, indicating a level within the top part of Zone N.21 (Kennett & Srinivasan, 1983). The assemblages from this part of the core contain Gr. (Gr.) multicamerata and Pu. praecursor indicating a level somewhat below the top of Zone N.21 (Kennett & Srinivasan, 1983; see also Fig. 2). Bolli Saunders (1985, fig. 10), however, show Gds. quadrilobatus fistulosus ranging from Zone N.19 into the base of Zone N.21 (effectively restricted to the middle Pliocene), barely overlapping with Gr. (Tr.) tosaensis. They also show the LAD of Gr. (Gr.) multicamerata occurring within Zone N.20, immediately before that of Gds. quadrilobatus fistulosus, with both taxa becoming extinct well below the FAD of Gr. (Tr.) truncatulinoides. Thus, according to Bolli & Saunders (1985) the assemblages from the three cores should be assigned to the basal part of Zone N.21. This conclusion is not consistent with the evidence of Kennett & Srinivasan (1983) nor with the data from this study. However, the presence of Gds. quadrilobatus fistulosus, Gr. (Gr.) multicamerata and Pu. praecursor within Zone N.22 in some of the cores studied, suggests that these taxa range into higher levels than considered by either Kennett & Srinivasan (1983) or Bolli & Saunders (1985).

Zone N.22. The subdivisions previously used for Zone N.22 (Bolli & Premoli Silva, 1973; Chaproniere, 1985b) have proved to be inadequate for this study, necessitating the use of new or redefined subzones. The base of Zone N.22 is defined

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НОГО			Gle	oborotalia tumida Zone	Lge. <i>Gr.tu.</i>	опе	Subzone	Gr.fi.		Hastigerina adamsi Subzone	Bo.a.		<i>Bolliella</i> <i>adamsi</i> Subzone	Bo.a.
	N.23	Gl.c. ⊥	Zone	Pulleniatina finalis		s Z	Globigerina bermudezi Subzone Globigerina calida calida Subzone	Gr.fl.	N.23	Pulleniatina obliquiloculata finalis Subzone Globigerina calida calida Subzone	Gr.fl.	N.23	Pulleniatina finalis Subzone Bolliella calida calida Subzone	Gr.fl. → Bo.c.
OCENE		- ,	tulinoides Z	Subzone	Pu.f. ⊥	ides truncatulinoide	Globorotalia hessi Subzone			Globorotalia crassaformis hessi			Bolliella praeadamsi Subzone Globorotalia crassaformis	<i>Bo.p.</i> <u></u> ⊥
P L E I S T	N.22		talia truncatu	<i>Globoquadrina</i> <i>dutertrei</i> Subzone		truncatulinoide		Gr.he. ⊥		Subzone	Gr.he. ⊥	N.22	<i>hessi</i> Subzone	Gr.he. ⊥
					Gr.to.		Globorotalia crassaformis			Globorotalia crassaformis			Globorotalia crassaformis viola Subzone	
		Gr.tr.	loboro	Globorotalia tosaensis		Globorotalia	<i>viola</i> Subzone	Gr.tr.		<i>viola</i> Subzone	Gr.tr.		Globigerinoides quadrilobatus fistulosus Subzone	Gd.fi.
N.21	→ Subzone	De.a.	trui	dulleniatina ncatulinoides . tosaensis Zone			N.21			N.21				

⁻ LAD Bo.a.-Bolliella adamsi Gr.fi.-Globorotalia(Gr.)fimbriatalge. Gr.tu.-large Gr.(Gr.)tumida tumida Gr.fl.-Gr.(Gr.)tumida flexuosa Gl.c.-Globigerínella calida calida Bo.c.-Bolliella calida calida Pu.f.-Pulleniatina finalis Bo.p.-Bolliella praeadamsi _____FAD Gr.he.-Gr.(Truncorotalia)crassaformis hessi Gd.fi.-Globigerinoides quadrilobatus fistulosus Gr.tr.-Gr.(Tr.)tosaensis

Figure 11. Planktic foraminiferal biostratigraphic zonation and boundary criteria for the Quaternary as discussed in this paper.

by the FAD of Gr. (Tr.) truncatulinoides, and the top by the FAD of Bo. calida calida (Blow, 1969).

De.a.-Dentoglobigerina altispira altispira

Globigerinoides quadrilobatus fistulosus Subzone. This subzone is defined by the interval between the FAD of Gr. (Tr.) truncatulinoides and the LAD of Gds. quadrilobatus fistulosus. This interval represents the lower part of Zone N.22, and is in part equivalent to the lower part of the Gr. (Tr.) crassaformis viola Subzone of Bolli & Premoli Silva (1973). The lower part of this subzone is present in core So16-13SL, but in this core the top of the subzone is marked by a disconformity.

The FAD of Gr. (Tr.) truncatulinoides, which occurs at 1.9 Ma and approximates the base of the Olduvai magnetic event (Berggren & others, 1985), marks the base of the subzone. The LAD of Gds. quadrilobatus fistulosus is given as 1.6 Ma, approximating the top of the Olduvai event (Berggren & others, 1985). Thus the Gds. quadrilobatus fistulosus Subzone had a duration of approximately 0.3 Ma.

Taxa having some biostratigraphic significance which range through this subzone are Gr. (Tr.) tosaensis and Gr. (Gr.) tumida flexuosa.

Taxa having some biostratigraphic significance which disappear within this subzone are De. altispira altispira, Gr. (Gr.) cultrata limbata, Gr. (Gr.) multicamerata and Pu. praecursor. Taxa having some biostratigraphic significance which appear within this subzone are Ga. (Go.) rubescens pink form and 'Tinophodella' evoluta.

Other significant biostratigraphic events: At least one sinistral coiling phase of Pu. obliquiloculata (L5 of Saito, 1976) is found in the studied sections. It is almost certain that another sinistral coiling phase for this species (L4) occurs within this subzone, but because of the disconformity within the lower part of the Pleistocene in the region, the position of this event has not been established by this study.

Globorotalia (Truncorotalia) crassaformis viola Subzone. This subzone was proposed by Bolli & Premoli Silva (1973), and was defined as that interval between the FAD of Gr. (Tr.) truncatulinoides and the FAD of Gr. (Tr.) crassaformis hessi. Because the newly defined Gds. quadrilobatus fistulosus Subzone is within the lower part of the Gr. (Tr.) crassaformis viola Subzone as originally defined, it is necessary to emend this definition. The base of the Gr. (Tr.) crassaformis viola Subzone is now defined by the LAD of Gds. quadrilobatus fistulosus, with the top being marked by the FAD of Gr. crassaformis hessi. Though the lower part of this subzone has not been recognised in the studied area because of the presence of a disconformity spanning part of the early Pleistocene, the top has been found in cores 51GC17 and 51GC22.

The LAD of Gds. quadrilobatus fistulosus has been dated at 1.6 Ma (Berggren & others, 1985). No published information is available for the FAD of Gr. (Tr.) crassaformis hessi, but in core 76PC06 this subspecies is present from the base of the core, and so this event must be older than oxygen isotope stage 12. However, in core 51GC22 both hessi and tosaensis are seen to overlap, and in core 51GC17 the two events are separated by 86 cm. Thus, the two events appear to be of similar age. The LAD of Gr. (Tr.) tosaensis has been given as 0.6 Ma by Berggren & others, (1985) giving a time interval of approximately 1.0 Ma for this subzone. Because few biostratigraphic events occur within this subzonal interval (Fig. 2), they are of little use for additional zonal subdivision. Only three occur at the very top (the FAD of Gr. (Ob.) bermudezi, and the LADs of Pu. praecursor and Ga. (Go.) decoraperta. Three intervals of sinistrally coiled populations of Pulleniatina seem to occur within the subzone (Saito, 1976), but these fall within the interval represented by a disconformity within the sections studied, indicating that only the top part of the subzone occurs within the study area.

Globorotalia (Truncorotalia) crassaformis hessi Subzone. This subzone was proposed by Bolli & Premoli Silva (1973) with the base defined by the FAD of Gr. (Tr.) crassaformis hessi and the top by the FAD of Bo. calida calida. Because the newly defined Bolliella praeadamsi Subzone occurs within the top part of this subzone as originally defined, it has been necessary to redefine the upper boundary on the FAD of Bo. praeadamsi. The Gr. (Tr.) crassaformis hessi Subzone succeeds the Gr. (Tr.) crassaformis viola Subzone in cores 51GC17 and 51GC22, but it is missing in core So16-13SL because a disconformity separates the Globigerinoides auadrilobatus fistulosus and Bolliella praeadamsi Subzones. The LAD of Gr. (Tr.) tosaensis occurs very close to the FAD of the subzonal marker and may be useful to indicate this interval in the absence of Gr. (Tr.) crassaformis hessi; indeed Kennett & Srinivasan (1983) have used the LAD of Gr. (Tr.) tosaensis to define the top of their Gr. (Tr.) truncatulinoides-Gr. (Tr.) tosaensis Zone. Berggren & others (1985) give the age of the LAD of Gr. (Tr.) tosaensis as 0.6 Ma which places it in oxygen isotope stage 16. The basal part of core 76PC06 falls within the Gr. (Tr.) crassaformis hessi Subzone and is older than oxygen isotope stage 10, evidence which does not dispute the conclusions of Berggren & others (1985). In core 76PC06 the top of the subzone falls below oxygen isotope stage 10, and is possibly within oxygen isotope stage 11 which is at approximately 0.36 Ma; thus, on the basis of this evidence, the subzone lasted approximately 0.24 Ma.

Bolliella praeadamsi Subzone. The base of this subzone is defined on the FAD of Bo. praeadamsi; the FAD of Pu. finalis occurs at the same time and so can be used as a supplementary definition. The top of the subzone is defined by the FAD of Bo. calida calida, an event which also marks the top of Zone N.22. As defined, the subzone falls within the upper part of the Gr. (Tr.) crassaformis hessi Subzone of Bolli & Premoli Silva (1973), making it necessary to emend that subzonal definition. The Bolliella praeadamsi Subzone is found in all cores studied, but the lower boundary occurs only in cores 51GC17, 51GC22, 51GC45 and 76PC06. As noted above, the base may fall within oxygen isotope stage 11 (0.36 Ma) in core 76PC06, and the top in oxygen isotope stage 8 (ca. 0.25 Ma). Bolli & Premoli Silva (1973) show the age of the FAD of Bo. calida calida to be 0.2 Ma, a date which falls within oxygen isotope stage 7, and is a similar level to that found in core 76PC06. Thus the interval of time occupied by the Bolliella praeadamsi Subzone ranges from 0.11 to 0.16 Ma.

Zone N.23. Bolli & Premoli Silva (1973) recognised three subzones within Zone N.23 in the Caribbean region, but I was

able to recognise only one of these (as the Globigerinella calida calida Subzone) near Wallis Island (Chaproniere, 1985b). Instead, I recognised two different subzones (the Pulleniatina finalis Subzone and the Hastigerina adamsi Subzone) (Fig. 2); the base of the Pulleniatina finalis Subzone was defined by the LAD of Gr. (Gr.) tumida flexuosa and the top by the FAD of Hastigerina adamsi (=Bolliella adamsi in this study). I have also been able to recognise the same sequence of subzones in the Coral Sea area. As noted above the FAD of Bo. calida calida occurs within oxygen isotope stage 7 in this region. Brönnimann & Resig (1971) noted the difficulty in separating the praecalida and calida morphotypes and so criticised the basis for Zone N.23.

Bolliella calida calida Subzone. The base of this subzone is defined by the FAD of *Bo. calida calida*, and the top by the LAD of *Gr. tumida flexuosa*, following Bolli & Premoli Silva (1973). This subzone has been recognised in all cores.

Two events occur within this subzone: the LAD of the pink form of *Gds. ruber*, and a change in the coiling direction for populations of *Gr. (Tr.) crassula* from dextral to sinistral. The coiling change takes place within oxygen isotope stage 7 in core 76PC06.

Pulleniatina finalis Subzone. Lamb & Beard (1972) described this subzone from the Caribbean region, and defined its base on the FAD of Pu. finalis and the top by the appearance of large specimens of Gr. (Gr.) tumida tumida. Because this sudden change in size of Gr. (Gr.) tumida tumida has not been observed in the Indo-Pacific region, I emended the definition of the subzone (Chaproniere, 1985b), defining the base on the LAD of Gr. (Gr.) tumida flexuosa and the top on the FAD of Bo. adamsi. This subzone can be recognised in all but one of the cores (51GC18), where the LAD of Gr. (Gr.) tumida flexuosa occurs below the FAD of Bo. calida calida. Over the upper part of its range this Gr. (Gr.) tumida flexuosa becomes rare and sporadic.

One biostratigraphic event occurs within this subzone, this being the change in coiling direction for populations of *Gr. (Tr.) crassula* from sinistral to dextral. This event occurs within a turbidite interval near the top of core 76PC06, making the correlation with the oxygen isotope stage chronology unreliable (Peerdeman, ANU, personal communication, 1990). Notwithstanding, this event must take place within the interval of isotope stages 1 to 4 in this core.

Bolliella adamsi Subzone. I defined the base of this subzone on the FAD of Bo. adamsi (Chaproniere, 1985b). This event occurs above the peak level of pink forms of Gds. ruber, an event which probably equates to that noted by Thompson & others (1979), who dated it at 120 000 BP. Rare specimens of this pink form are present in Holocene sediments, both in the northern Australian region, as well as further to the east (Chaproniere, 1985b), which limits its biostratigraphic usefulness. However, in all cores there is a sudden reduction in population size, and this event appears to be biostratigraphically useful. This subzone is present in all cores. In core 76PC06 the FAD of Bo. adamsi occurs only in the disturbed upper part, making correlation with the oxygen isotope stratigraphy inappropriate. However, this event must take place within either isotope stages 1 or 2, making it a possible marker for the Holocene.

The LAD of *Gr.* (*Tr.*) crassaformis hessi is the only event to take place within this subzone. Towards the end of its range this subspecies becomes very rare, reducing its potential as a zonal indicator.

Comparison with previous zonations

Figure 11 illustrates a comparison between the zonal scheme proposed in this paper and those of other papers and shows the relationship of events to the oxygen isotope stratigraphic scheme established in the Coral Sea area. The region studied for this paper has been within the tropical belt throughout the Pleistocene, and so has been minimally affected by the temperature changes which occurred over this time interval. All other studies are based on areas near the subtropical/warm temperate transition, and their zonal schemes consequently reflect the faunal changes related to fluctuations in water temperature. In addition, a degree of provincialism can be recognised in the faunas from the different areas, making it necessary for new subzones to be established for the Coral Sea

Environmental controls

The association Globigerinita glutinata, Gds. conglobatus, Gds. ruber, Gds. sacculifer, Gr. (Tr.) crassaformis, Gr. (Gr.) tumida, N. dutertrei and Pu. obliquiloculata is generally considered to be tropical (Bé & Tolderlund, 1971), being typical of waters north of 15°S in the western Atlantic Ocean (van Leeuwen, 1989). In these associations N. dutertrei is always dominantly dextrally coiled. Further, Gr. (Tr.) truncatulinoides is dominated by dextrally coiled individuals in subtropical and tropical seas (Healy-Williams, 1983). However, for many species temperature does not appear to be the main controlling factor; the physical and chemical properties of the water mass are more important (Bé, 1977). For example, Thiede (1971) found that salinity appears to have controlled the coiling direction of Gr. (Tr.) truncatulinoides during the north Atlantic Quaternary, with sinistrally coiled populations being typical of interglacial sediments. Thiede (1971) concluded that changes in salinity due to expansion of the central north Atlantic water mass controlled coiling direction. Bé & Tolderlund (1971) also showed that there was a sharp boundary between sinistral and dextrally coiled populations at 25°S in the Indian Ocean, where there appears to be no major change in physical oceanographic characteristics. More recently, van Leeuwen (1989) and Auras-Schudnagies & others (1989) have shown that nutrient levels control the distribution of some species; for example Globigerina (Ga.) bulloides occurs in nutrient-rich areas such as upwellings, whereas Globigerinoides sacculifer and Gds. ruber are typical of oligotrophic areas.

The Pleistocene planktic foraminiferal faunas from the cores show a marked stability, with the assemblages dominated by oligotrophic, warm water species such as Gds. quadrilobatus sacculifer, Gds. ruber and Gr. (Gr.) cultrata. Pulleniatina, Neogloboquadrina (both dextrally coiled), Sphaeroidinella dehiscens and Gr. (Gr.) tumida, though never abundant, are universally present. Thus, on the faunal evidence, warm oligotrophic conditions similar to those today have been stable in the region since at least the middle Pleistocene.

Conclusions

The study of seven cores from the Coral Sea off northeastern Australia has permitted the assessment of 27 planktic foraminiferal biostratigraphic events from the latest Pliocene, basal Pleistocene and mid to late Pleistocene; most of the early Pleistocene could not be studied because of a hiatus in the one core which penetrated through the Pleistocene into the top of the Pliocene. Some of these events have been used to erect a formal zonal and subzonal scheme for the region, building upon studies by Blow (1969), Bolli & Premoli Silva (1973),

Rögl (1974) and Chaproniere (1985b). Following my earlier study (Chaproniere, 1985b), Zones N.22 and N.23 of Blow (1969) have been used, and the modified subzonal scheme of Bolli & Premoli Silva (1973), used by Chaproniere (1985b) has been modified slightly, with the addition of the new Globigerinoides quadrilobatus fistulosus and the Bolliella praeadamsi Subzones. This has made necessary the redefinition of the Globorotalia (Truncorotalia) crassaformis viola and the Globorotalia (Truncorotalia) crassaformis hessi Subzones of Bolli & Premoli Silva (1973). There is little faunal evidence for changes in palaeoceanographic conditions being related to the faunal changes observed in the cores, although there is strong evidence of reworking at some levels within some of the cores (such as late Miocene Zone N.17 in core 51GC17), which may have been related to sea-level fluctuations. The assemblages from the cores show a marked stability, with the faunas being dominated by spinose, oligotrophic taxa. Temperate water forms are either very rare or absent, whereas tropical and subtropical species are dominant.

A single new species, Bolliella praeadamsi, has been described because of its biostratigraphic utility. This species is an evolutionary intermediate between Bo. calida praecalida and Bo. adamsi.

Acknowledgements

I wish to thank the scientists and crews of the RV Sonne and Rig Seismic, without whose help I would not have had the samples used in this study. I am very grateful to Frank Peerdeman (ANU) for access to the results of a reconnaissance study of oxygen isotopes for core 76PC06. I also thank Frank Peerdeman, Peter Davies (BMR) and Samir Shafik (BMR) for critically reading various drafts of this paper.

Systematic description

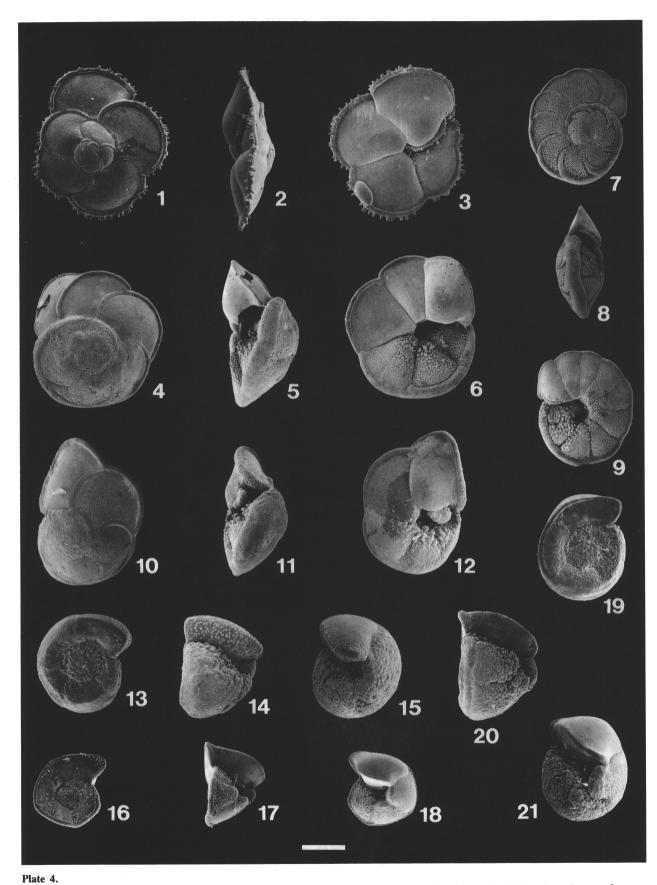
Family Globigerinidae Subfamily Hastigerininae

The following discussion is restricted to the hastigerines in order to describe a new species which has biostratigraphic importance for the region. The taxonomic nomenclature used herein is mainly in accordance with Kennett & Srinivasan (1983), Saito & others (1981) and Loeblich & Tappan (1988).

Genus Bolliella Banner & Blow, 1959

Type species, Hastigerina (Bolliella) adamsi Banner & Blow. 1959; by original designation.

Banner & Blow (1959) established Bolliella as a subgenus of Hastigerina on the basis that the chambers were radially elongate in the adult. Included in the concept of Hastigerina was Globigerinella following Bolli & others (1957). Banner & Blow (1960) maintained their previous concept of Hastigerina and considered Globigerinella aequilateralis a junior synonym of H. siphonifera. Loeblich & Tappan (1964) elevated Bolliella to full generic status. Bé (1967, 1969) believed that spine shape was an important taxonomic character and used it to separate Hastigerina from Globigerinella; both were shown to have triradiate spines, but Globigerinella differed in having simple spines also. This separation has been supported by Loeblich & Tappan (1988) who considered that Bolliella had an apertural lip which was absent in Globigerinella. Saito & others (1981), without giving reasons, believed Bolliella to be a junior synonym for Globigerinella and referred adamsi to that genus. Kroon (1988) noted that in the 125 to 250 µm size fraction, specimens of adamsi, calida and siphonifera were difficult to separate.



FIALE 4.

1-3, Globorotalia (Globorotalia) cultrata fimbriata, CPC30240, sample 81640043, So16-13SL-260. 4-6, Globorotalia (Globorotalia) cultrata neoflexuosa, CPC30241, sample 51GC45-121. 7-9, Globorotalia (Globorotalia) multicamerata, CPC30242, sample 81640043, So16-13SL-260. 10-12, Globorotalia (Globorotalia) tumida flexuosa, CPC30243, sample 76PC06-190. 13-15, Globorotalia (Truncorotalia) tosaensis, CPC30244, sample 51GC17-328. 16-18, Globorotalia (Truncorotalia) truncatulinoides truncatulinoides, CPC30245, sample 51GC22-040. 19-21, Globorotalia (Truncorotalia) truncatulinoides pachytheca, CPC30246, sample 51GC22-040.

Scale bar figs 1-12, 16-21, 300 μm; 13-15, 200 μm.

Hastigerina differs from both Globigerinella and Bolliella in having only triradiate spines, whilst the two other genera have both rounded and triradiate spines. The evidence presented by Saito & others (1976) showing that the surface texture of Bolliella and Globigerinella to be very similar lead them to consider that both Bo. adamsi and Gl. siphonifera were closely related, with Bo. calida probably being the ancestral form of Bo. adamsi. Though both spine shape and surface texture are important morphologic features, the presence of an apertural lip also appears important (Loeblich & Tappan, 1988). The type species of Globigerinella, Gl. aequilateralis, lacks an apertural lip (Saito & others, 1981) a feature observed in this study (Plate 1, figs 14, 17), whilst the adamsi, calida and praeadamsi morphotypes invariably possess a thin apertural lip. Even though a number of workers believe that Bolliella should be considered a junior synonym of Globigerinella, I believe the chamber elongation and the presence of an apertural lip in Bolliella to be sufficient and easily recognisable morphologic characters for distinguishing Bolliella from Globigerinella. In addition I believe that the adamsi, calida, praecalida and praeadamsi morphotypes share enough characteristics for them all to be referred to Bolliella. Furthermore, on phylogenetic grounds it is best to maintain this separation.

Saito & others (1976) considered that Bo. calida was the most probable ancestor of Bo. adamsi, and noted that the only characters separating the two species were the chamber elongation and the planispiral coiling of Bo. adamsi.

Bolliella praeadamsi new species Plate 1, figs 1-6, 19-21; Plate 2, figs 1-9

Types. Holotype and 5 paratypes from sample 81640021, So16-12SL-075, and are filed under numbers CPC30216-30218 and CPC30224-30226 in the Commonwealth Palaeontology Collection, BMR, Canberra.

Diagnosis. A hastigerine with the final 1 to 4 chambers arranged planispirally, and which may or may not show some radial elongation; the earlier part of the final whorl is clearly trochospiral.

Description. Test medium to large, with 5 or 6 chambers in the final whorl. Initial stage a low trochospire, with the last 1 to 4 chambers planispirally arranged and well separated from the previous whorl. Chambers initially spherical and globular becoming ovate; may finally be radially elongate in large specimens. Chambers increase rapidly in size throughout ontogeny, each separated from the previous chamber by a depressed radial suture. Aperture umbilical-extra-umbilical and a low asymmetrical arch on the earlier chambers of the final whorl, becoming a high and wide symmetrical arch with a distinct thin lip. Wall calcareous, spinose, densely perforate with large circular pores, with randomly distributed, rounded and raised spine bases. Spines round becoming triradiate.

Remarks. In side view this species is very similar to Bo. calida sensu lato in having a narrow test, but differs by having 1-4 chambers arranged in a planispire, clearly separated from the previous whorl. In addition, some of the final chambers may show some degree of radial elongation. Bo. praeadamsi differs from Bo. adamsi in having rounded, subglobular chambers, rather than the pointed distal chamber apex of Bo. adamsi. It differs from Gl. aequilateralis (1) in having only the last few chambers of the test arranged planispirally, with a distinct trochospiral part of the final whorl as viewed from the spiral side, (2) by having more chambers in the final whorl, the earlier part of the test being somewhat narrower in side view, (3) by having the last few chambers separated from the

previous whorl, and (4) by having a distinct apertural lip. The chambers of Gl. aequilateralis are more globular and expand more rapidly in diameter as the test increases in size, leading to a thicker test as seen in side view. Saito & others (1981) show that the test wall of Bo. calida is very similar to that of Bo. adamsi, a feature not found in this study. The wall texture of both Bo. adamsi and Bo. praeadamsi is identical, but it has distinctly larger diameter pores than those of Bo. calida. This may be an environmental factor, although specimens illustrated by Saito & others (1981) were collected from the southwestern equatorial Pacific, not far from the present study area.

This species has almost certainly been recorded either as Gl. aequilateralis (= Gl. siphonifera) or as Bo. adamsi, with which it has many features in common. It is readily differentiated from Gl. aequilateralis in side view. It is very similar to both Bo. calida and Bo. adamsi in side view in its slow rate of chamber expansion, resulting in a lower trochospire than that of Gl. aequilateralis.

Stratigraphic Range. Middle Pleistocene, Zone N.22 (Bo. praeadamsi Subzone) to Recent (oxygen isotope stages 1 to 11 or 12).

Phylogeny. Two phylogenies for the hastigerines have been proposed. Blow (1969) traced the phylogenetic origins of this group to Gl. obesa. He believed that this morphotype graded into the planispiral forms Gl. praesiphonifera (within Zone N.7), Gl. aequilateralis (= siphonifera) in Zone N.12, and Bo. adamsi within the Pleistocene (Banner & Blow, 1960). Blow (1969) considered that Bo. calida praecalida had its origins within the Globigerina praebulloides group and gave rise to Bo. calida calida, which is thus unrelated to Hastigerina. H. pelagica was considered to have had an unknown ancestry (Banner & Blow, 1960). Kennett & Srinivasan (1983, text fig. 26) proposed a phylogenetic scheme for the group, with the lineage (Gl.obesa-Gl. praesiphonifera-Gl. aequilateralis) as in Banner & Blow (1960). However, H. pelagica and Bo. calida (including both the praecalida and calida morphotypes) were both derived from Gl. aequilateralis (within Zones N.17A and N.19, respectively). Srinivasan & Kennett (1975) believed that Bolliella evolved from Globigerinella.

As noted above, the presence of an apertural lip and the arrangement of the initial coil of Bo. calida praecalida, Bo. calida calida, Bo. praeadamsi and Bo. adamsi distinguishes the group from other spinose forms such as Globigerina bulloides, Gl. aequilateralis and Hastigerina pelagica. On the basis of wall structure (size and pattern of pores, characteristics of spines and spine bases), the Bo. calida group and Bolliella are close to Gl. aequilateralis, but distinct from Hastigerina. The FAD of Bo. praeadamsi occurs before the FAD of Bo. calida calida, indicating that the latter cannot be the ancestral form. As noted above, the early ontogenetic stage of Bo. praeadamsi is very similar to Bo. calida praecalida, and this appears to be the preferred ancestral form of Bo. praeadamsi.

References

Auras-Schudnagies, A., Kroon, D., Ganssen. G.M., Hemleben, C. & Van Hinte, J.E., 1989 — Biogeographic evidence from planktic foraminifers and pteropods for Red Sea anti-monsoonal surface currents. *Deep-Sea Research*, 36, 1515–1534.
Banner, F.T. & Blow, W.H., 1959 — The classification and stratigra-

phical distribution of the Globigerinaceae. Part 1. Palaeontology, 2, 1-27

Banner, F.T. & Blow, W.H., 1960 — The taxonomy, morphology and affinities of the genera included in the subfamily Hastgerininae. Micropaleontology, 6, 19-31.

- Banner, F.T. & Blow, W.H., 1967 The origin, evolution and taxonomy of the foraminiferal genus Pulleniatina Cushman, 1927. Micropaleontology, 13, 133-162.
- Bé, A.W.H., 1967 Foraminifera. Families: Globigerinidae and Globorotaliidae. Fiches d'identification du zooplancton. Conseil Permanent International pour l'Exploration de la Mer Charlottenlund Slot, Danemark, Sheet 108, 1-6.
- Bé, A.W.H., 1969 Microstructural evidence of the close affinity of Globigerinella Cushman to Hastigerina Thomson. In Brönnimann, P. & Renz, H.H. (editors), Proceedings of the First International Conference on Planktonic Microfossils, I, 90-91.
- Bé, A.W.H., 1977 An ecologic, zoogeographic and taxonomic review of Recent planktonic foraminifera. In A.T.S. Ramsay (editor), Oceanic micropalaeontology, 1-100. Academic Press, London.
- Bé, A.W.H. & Tolderlund, D.S., 1971 Distribution and ecology of living planktonic foraminifera in surface waters of the Atlantic and Indian Oceans. In Funnell, B.M. & Riedel, W.R. (editors), Micropaleontology of oceans. Cambridge University Press, London, 105-149
- Beckmann, J.P., 1971 The foraminifera of Sites 68-75. In Tracey, J.I. jnr. & others, Initial Reports of the Deep Sea Drilling Program, 8, 713-725.
- Belford, D.J., 1988 Late Tertiary and Quaternary foraminifera and pal aeobathymetry of dredge and core samples from the New Ireland Basin (Cruise L7-84-SP). In Marlow, M.S., Dadiswan, S. & Exon, N.F. (editors), Geology and offshore resources of Pacific island arcs - New Ireland and Manus region, Papua New Guinea. Circum-Pacific Council for Energy and Mineral Resources Earth Science Series, 9, 65-89. Circum-Pacific Council for Energy and Mineral Resources, Houston, Texas.
- Berggren, W.A. & Boersma, A., 1969 Late Pleistocene and Holocene planktonic foraminifera from the Red Sea. In Degans, E.T. & Ross, D.A. (editors), Hot brines and recent heavy metal deposits in the Red Sea. Springer-Verlag, New York, 282-298.
- Berggren, W.A., Burkle, L.H., Cita, M.B., Cooke, H.B.S., Funnell, B.M., Gartner, S., Hays, J.D., Kennett, J.P., Opdyke, N.D., Pastouret, L., Shackleton, N.J. & Takayanagi, Y., 1980 — Towards a Quaternary time scale. Quaternary Research, 13, 277-
- Berggren, W.A., Kent, D.V. & van Couvering, J.A., 1985 The Neogene: Part 2. Neogene geochronology and chronostratigraphy. In Snelling, N.J. (editor), The chronology of the geological record. The Geological Society of London, Memoir 10, 211-250.
- Blow, W.H., 1969 Late Middle Eocene to Recent planktonic foraminiferal biostratigraphy. In P.Brönnimann & H.H. Renz (editors), Proceedings of the First International Conference on Planktonic Microfossils, I, 199-421, E.J. Brill, Leiden.
- Bolli, H.M., 1966 The planktonic foraminifera in well Bodjonegoro-1 of Java. Eclogae Geologicae Helvetiae, 59, 449-465.
- Bolli, H.M., Loeblich, A.R., jnr. & Tappan, H., 1957 Planktonic foraminiferal families Hantkeninidae, Orbulinidae, Globorotalidae and Globotruncanidae. *In* Loeblich, A.R. jnr., Tappan, H., Beckmann, J.P., Bolli, H.M., Gallitelli, E.M. & Troelsen, J.C., Studies in Foraminifera. United States National Museum, Bulletin 215, 1-50. Smithsonian Institution, Washington.
- Bolli, H.M. & Premoli Silva, I., 1973 Oligocene to Recent planktonic foraminifera and stratigraphy of the Leg 15 sites in the Caribbean Sea. In Edgar, N.T., Saunders, J.B. & others, 1973 Initial Reports of the Deep Sea Drilling Project, 15, 475-497. U.S. Government Printing Office, Washington.
- Bolli, H.M., & Saunders, J.B., 1985 Oligocene to Holocene low latitude planktic foraminifera. In Bolli, H.M., Saunders, J.B. & Perch-Nielson, K. (editors), Plankton stratigraphy. Cambridge University Press, Cambridge, 155-262.
- Brönnimann, P. & Resig, J.M., 1971 A Neogene globigerinacean biochronologic time-scale of the Southwestern Pacific. Initial Reports of the Deep Sea Drilling Project, 7, 1235-1469. U.S. Government Printing Office, Washington.
- Chaproniere, G.C.H., 1985a Late Neogene and Quaternary planktonic foraminiferal biostratigraphy and paleobathymetry of dredge samples from the southern Tongan Platform (Cruise L5-82-SP). In Scholl, D.W. & Vallier, T.L. (editors), Geology and offshore resources of Pacific island arcs — Tonga region. Circum-Pacific Council for Energy and Mineral Resources Earth Science Series, 2, 131-140. Circum-Pacific Council for Energy and Mineral Resources, Houston.

- Chaproniere, G.C.H., 1985b Late Tertiary and Quaternary foraminiferal biostratigraphy and paleobathymetry of cores and dredge samples from Cruise KK820316 Leg 2. In Brocher, T.M. (editor), Geological investigations of the northern Melanesian Borderland. Circum-Pacific Council for Energy and Mineral Resources Earth Science Series, 3, 103-122. Circum-Pacific Council for Energy and Mineral Resource, Houston, Texas.
- Chaproniere, G.C.H., in press Late Eocene to Pleistocene foraminiferal biostratigraphy and paleobathymetry of dredge samples from the southern Tonga Platform (Cruise L3-84-SP). In Ballance, P.F., Herzer, R.H. & Vallier, T.L. (editors), Contributions to the geology of the Tonga and Lau regions of the Southwest Pacific. Circum-Pacific Council for Energy & Mineral Resources, Earth Science Series, Houston, Texas.
- Dowsett, H.J., 1989 Application of the graphic correlation method to Pliocene marine sequences. Marine Micropaleontology, 14, 3-32.
- Ericson, D.B. & Wollin, G., 1968 Pleistocene climates and chronology in deep-sea sediments. Science, 162, 1227-1234.
- Fordham, B.G., 1986 Miocene-Pleistocene planktic foraminifers from DSDP Sites 208 and 77, and phylogeny and classification of Cenozoic species. Evolutionary Monographs, University of Chicago, 6, 1-200.
- Healy-Williams, N., 1983 Fourier shape analysis of Globorotalia truncatulinoides from late Quaternary sediments in the southern Indian Ocean. Marine Micropaleontology, 8, 1-15.
- Herman, Y., 1968 Evidence of climatic changes in Red Sea cores: means of correlation of Quaternary successions. 8th Congress of the International Association of Quaternary Research, 325-348.
- Hills, S.J. & Thierstein, H.R., 1989 Plio-Pleistocene calcareous plankton biochronology. Marine Micropaleontology, 14, 67-96.
- Jenkins, D.G., 1967 Planktonic foraminiferal zones and new taxa from the Lower Miocene to the Pleistocene of New Zealand, New Zealand Journal of Geology and Geophysics, 10, 1064-1078.
- Jenkins, D.G. & Orr, W.N., 1972 Planktonic foraminiferal biostratigraphy of the eastern Equatorial Pacific. DSDP Leg 9. In Hays, J.D. & others, Initial Reports of the Deep Sea Drilling Project, 9, 1059-1193. U.S. Government Printing Office, Washin-
- Kennett, J.P., 1976 Phenotypic variation in some Recent and Late Cenozoic planktonic foraminifera. In Hedley, R.H. & Adams, C.G. (editors), Foraminifera, 2, 111-169. Academic Press, Lon-
- Kennett, J.P. & Huddlestun, P., 1972 Abrupt climatic change at 90,000 years B.P.: faunal evidence from Gulf of Mexico cores. Quaternary Research, 2, 384-395.
- Kennett, J.P. & Srinivasan, M.S., 1983 Neogene planktonic foraminifera. A phylogenetic atlas. Hutchinson Ross, Stroudsburg, Pennsylvania, 1-265.
- Kroon, D., 1988 Distribution of extant planktic foraminiferal assemblages in Red Sea and northern Indian Ocean surface waters. In Brummer, G.J.A. & Kroon, D., Planktonic foraminifers as tracers of ocean-climate history. VU Uitgeverij, Amsterdam, 229-267.
- Lamb, J.L. & Beard, J.H., 1972 Late Neogene planktonic foraminifers in the Caribbean, Gulf of Mexico, and Italian stratotypes. The University of Kansas Paleontological Contributions, Article 57, 1-67.
- Locke, S. & Thunell, R.C., 1988 Palaeoceanographic record of the last glacial/interglacial cycle in the Red Sea and Gulf of Aden. Palaeoceanography, Palaeoclimatology, Palaeoecology, 63, 163-187
- Loeblich, A.R., jnr. & Tappan, H., 1964 On 'Hastigerina digitata Rhumbler, 1911' — Comment. Micropaleontology, 10, 494-495.
- Loeblich, A.R. & Tappan, H., 1988 Foraminiferal genera and their classification. Van Nostrand Reinhold, New York, 970 pp.
- Parker, F.L., 1967 Late Tertiary biostratigraphy (planktonic foraminifera) of tropical Indo-Pacific deep-sea cores. Bulletins of American Paleontology, 52, 115-208.
- Poag, C.W. & Valentine, P.C., 1976 Biostratigraphy and ecostratigraphy of the Pleistocene basin Texas-Louisiana continental shelf. Transactions of the Gulf Coast Association of Geological Societies, 26, 185-256.
- Reiss, Z., Luz, B., Almogi-Labin, A., Halicz, E., Winter, A. & Wolf, M., 1980 — Late Quaternary paleoceanography of the Gulf of Aqaba (Elat), Red Sea. Quaternary Research, 14, 294-308.

- Rio, D., Sprovieri, R., Stefano, E.D. & Raffi, I., 1984a Globorotalia truncatulinoides (d'Orbigny) in the Mediterranean upper Pliocene geologic record. Micropaleontology, 30, 121–137.
- Rio, D., Sprovieri, R. & Raffi, I., 1984b Calcareous plankton biostratigraphy and biochronology of the Pliocene-Lower Pleistocene succession of the Capo Rossello area, Sicily. *Marine Micropaleontology*, 9, 135–180.
- Rögl, F., 1974 The evolution of the Globorotalia truncatulinoides and Globorotalia crassaformis group in the Pliocene and Pleistocene of the Timor Trough, DSDP Leg 27, Site 262. In Veevers, J.J., Heirtzler, J.R. & others, Initial Reports of the Deep Sea Drilling Project, 27, 743-767. U.S. Government Printing Office, Washington.
- Rögl, F. & Bolli, H.M., 1973 Holocene to Pleistocene planktonic foraminifera of Leg 15, Site 147 (Cariaco Basin (Trench), Caribbean Sea) and their climatic interpretation. In Edgar, N.T., Saunders, J.B. & others, Initial Reports of the Deep Sea Drilling Project, 15, 553-616. U.S. Government Printing Office, Washington.
- Saito, T., 1976 Geologic significance of coiling direction in the planktonic foraminifera *Pulleniatina*. *Geology*, 4, 305-309.
- Saito, T., Burkle, L.H. & Hays, J.D., 1975 Late Miocene to Pleistocene biostratigraphy of equatorial Pacific sediments. In Saito, T. & Burkle, L.H. (editors), Late Neogene epoch boundaries. Micropaleontology Press, New York, 226-244.
- Saito, T., Thompson, P.R. & Breger, D., 1976 Skeletal ultrastructure of some elongate-chambered foraminifera and related species. In Takayanagi, Y. & Saito, T. (editors), Progress in micropaleontology. Micropaleontology Press, New York, 278-304.
- Saito, T., Thompson, P.R. & Breger, D., 1981 Systematic index of Recent and Pleistocene planktonic Foraminifera. *University of Tokyo Press*, *Tokyo*, 1–190.
- Shackleton, N.J. & Opdyke, N.D., 1976 Oxygen isotope and paleomagnetic stratigraphy of Pacific core V28-239: Late Pliocene to latest Pleistocene. *In* Cline, R.M. & Hays, J.D. (editors), Investigations of Late Quaternary paleoceanography and paleo-

- climatology. Memoir of the Geological Society of America, 145, 449-464.
- Srinivasan, M.S. & Kennett, J.P., 1975 The status of Bolliella, Beella, Protentella and related planktonic foraminifera based on surface ultrastructure. Journal of Foraminiferal Research, 5, 155– 165.
- Thiede, J., 1971 Variations in coiling ratios of Holocene planktonic foraminifera. *Deep-Sea Research*, 18, 823–831.
- Thompson, P.R., B , A.W.H., Duplessy, J.-C. & Shackleton, N.J., 1979 — Disappearance of pink-pigmented Globigerinoides ruber at 120,000 years BP in the Indian and Pacific Oceans. Nature, 280, 554–558.
- Thompson, P.R. & Sciarrillo, J.R., 1978 Planktonic foraminiferal biostratigraphy in the equatorial Pacific. *Nature*, 276, 29–33.
- Thunell, R.C., 1984 Pleistocene planktonic foraminiferal biostratigraphy and paleoclimatology of the Gulf of Mexico. In N. Healy-Williams (editor), Principles of Pleistocene stratigraphy applied to the Gulf of Mexico. D. Reidel Publishing Co., Boston, 25-64.
- Thunell, R.C., Williams, D.F. & Kennett, J.P., 1977 Late Quaternary paleoclimatology, stratigraphy, and sapropel history in eastern Mediterranean deep-sea sediments. *Marine Micropaleon-tology*, 2, 371–388.
- Tracey, J.I. jnr., Sutton, G.H., Nesteroff, W.D., Galehouse, J., von der Borch, C.C., Moore, T.C., Haq, B.U. & Beckmann, J.P., 1971a Site 71. In Tracey, J.I. jnr. & others, Initial Reports of the Deep Sea Drilling Project, 8, 285–458. U.S. Government Printing Office, Washington.
- Tracey, J.I. jnr., Sutton, G.H., Nesteroff, W.D., Galehouse, J., von der Borch, C.C., Moore, T.C., Haq, B.U. & Beckmann, J.P., 1971b Site 72. In Tracey, J.I. jnr. & others, Initial Reports of the Deep Sea Drilling Project, 8, 459–533. U.S. Government Printing Office, Washington.
- Van Leeuwen, R.J.W., 1989 Sea-floor distribution and late Quaternary faunal patterns of planktonic and benthic foraminifers in the Angola Basin. *Utrecht Micropaleontology Bulletins*, 38, 1–287.